



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : C12N 15/29, C12Q 1/68, G01N 33/53		A1	(11) International Publication Number: WO 99/02689
			(43) International Publication Date: 21 January 1999 (21.01.99)
(21) International Application Number: PCT/US98/13992 (22) International Filing Date: 7 July 1998 (07.07.98) (30) Priority Data: 08/893,401 11 July 1997 (11.07.97) US (71) Applicant (for all designated States except US): E.I. DU PONT DE NEMOURS AND COMPANY [US/US]; 1007 Market Street, Wilmington, DE 19898 (US). (72) Inventors; and (75) Inventors/Applicants (for US only): ODELL, Joan, Tellefsen [US/US]; 17 Monitor Place, Unionville, PA 19375 (US). ALLEN, Stephen, M. [US/US]; 12 Stanton Avenue, West Chester, PA 19382 (US). (74) Agent: MAJARIAN, William, R.; E.I. du Pont de Nemours and Company, Legal Patent Records Center, 1007 Market Street, Wilmington, DE 19898 (US).		(81) Designated States: AL, AM, AU, AZ, BA, BB, BG, BR, BY, CA, CN, CU, CZ, EE, GE, GW, HR, HU, ID, IL, IS, JP, KG, KP, KR, KZ, LC, LK, LR, LT, LV, MD, MG, MK, MN, MX, NO, NZ, PL, RO, RU, SG, SI, SK, SL, TJ, TM, TR, TT, UA, US, UZ, VN, YU, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG). Published With international search report.	
(54) Title: PLANT SUG1 HOMOLOGS			
(57) Abstract			
<p>This invention relates to an isolated nucleic acid fragment encoding a plant SUG1 protein, a homolog of the mouse SUG1 protein. The invention also relates to the construction of a chimeric gene encoding all or a portion of the plant SUG1 protein, in sense or antisense orientation, wherein expression of the chimeric gene results in production of altered levels of the plant SUG1 protein in a transformed host cell.</p>			
1	MATVAMDISKPTPAASGDEAAAAAKGRSGGGGEGLRQYYLQHIHDLQLQIRQKTHNLRL		
2	MALVGVLELKHAAEGVFEANCSA...KPTKQEGELRHYYSLNIHEHQLLRQKTHNLRL		
3	MALDGPPEMELEEG.....KAGSGLRQYYLSKIEELQLIVNDKSNLRL		
4	MATVAMDISKPTPAASGDEAAAAAKGRSGGGGEGLRQYYLQHIHDLQLQIRKTHNLRL		
1	EAQRNDLNSRVRLREELQLLQEPGSYVGEVVKVMGSKVLVVRPEGKYVVDIDKSIDI		
2	EAQRNDLNSRVRLREELQLLQEPGSYVGEVVKVMGSKVLVVRPEGKYVVDIDKSIDI		
3	QAQRNELNAKVRLLREELQLLQEPGSYVGEVVRAMDKCKVLVVRPEGKYVVDIDKSIDI		
4	EAQRNDLNSRVRLREELQLLQEPGSYVGEVVKVMGSKVLVVRPEGKYVVDIDKSIDI		
1	TKITPSTRVALRNDSYMLHLILPSKVDPLVNLKVEKVPDSTYDMIGGLDQIKIKEIVI		
2	TKITPSTRVALRNDSYMLHLILPSKVDPLVNLKVEKVPDSTYDMIGGLDQIKIKEIVI		
3	NDVTPNCRVALRNDSYMLHLILPSKVDPLVNLKVEKVPDSTYDMIGGLDQIKIKEIVI		
4		
1	ELPIKHPFELFESLGIAQPKGVLLYGPPGTGKTLARAVAHHTDCTFIRVSGSELVQKYIG		
2	ELPIKHPFELFESLGIAQPKGVLLYGPPGTGKTLARAVAHHTDCTFIRVSGSELVQKYIG		
3	ELPVKHPFELFESLGIAQPKGVLLYGPPGTGKTLARAVAHHTDCTFIRVSGSELVQKYIG		
4		
1	EGSRMVRELFVMAREHAPSIIIFMDEIDSIGSGARMESGTGNGDSEVQRTMLELLNQLDGE		
2	EGSRMVRELFVMAREHAPSIIIFMDEIDSIGSGARMESGTGNGDSEVQRTMLELLNQLDGE		
3	EGSRMVRELFVMAREHAPSIIIFMDEIDSIGSGARMESGTGNGDSEVQRTMLELLNQLDGE		
4		
1	ASNKIKVLMATNRIDILDQALLAPGRIDRKIEFPNPNEDSRFDILKHSRMMNLMRGIDL		
2	ASNKIKVLMATNRIDILDQALLAPGRIDRKIEFPNPNEDSRFDILKHSRMMNLMRGIDL		
3	ATRNKIVLMATNRIDILDQALLAPGRIDRKIEFPNPNEDSRFDILKHSRMMNLMRGIDL		
4		
1	KKIAEKONGASGAELKAVCTEAGMFALRERRVHVTOEDFEMAVAKVMKDKTEKMSLRKL		
2	KKIAEKONGASGAELKAVCTEAGMFALRERRVHVTOEDFEMAVAKVMKDKTEKMSLRKL		
3	KKIAELMPGASGAELKAVCTEAGMFALRERRVHVTOEDFEMAVAKVMKDKTEKMSLRKL		
4		
1	WK		
2	WK		
3	WK		
4	..		

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AL	Albania	ES	Spain	LS	Lesotho	SI	Slovenia
AM	Armenia	FI	Finland	LT	Lithuania	SK	Slovakia
AT	Austria	FR	France	LU	Luxembourg	SN	Senegal
AU	Australia	GA	Gabon	LV	Larvia	SZ	Swaziland
AZ	Azerbaijan	GB	United Kingdom	MC	Monaco	TD	Chad
BA	Bosnia and Herzegovina	GE	Georgia	MD	Republic of Moldova	TG	Togo
BB	Barbados	GH	Ghana	MG	Madagascar	TJ	Tajikistan
BE	Belgium	GN	Guinea	MK	The former Yugoslav Republic of Macedonia	TM	Turkmenistan
BF	Burkina Faso	GR	Greece			TR	Turkey
BG	Bulgaria	HU	Hungary	ML	Mali	TT	Trinidad and Tobago
BJ	Benin	IE	Ireland	MN	Mongolia	UA	Ukraine
BR	Brazil	IL	Israel	MR	Mauritania	UG	Uganda
BY	Belarus	IS	Iceland	MW	Malawi	US	United States of America
CA	Canada	IT	Italy	MX	Mexico	UZ	Uzbekistan
CF	Central African Republic	JP	Japan	NE	Niger	VN	Viet Nam
CG	Congo	KE	Kenya	NL	Netherlands	YU	Yugoslavia
CH	Switzerland	KG	Kyrgyzstan	NO	Norway	ZW	Zimbabwe
CI	Côte d'Ivoire	KP	Democratic People's Republic of Korea	NZ	New Zealand		
CM	Cameroon		Republic of Korea	PL	Poland		
CN	China	KR	Republic of Korea	PT	Portugal		
CU	Cuba	KZ	Kazakhstan	RO	Romania		
CZ	Czech Republic	LC	Saint Lucia	RU	Russian Federation		
DE	Germany	LI	Liechtenstein	SD	Sudan		
DK	Denmark	LK	Sri Lanka	SE	Sweden		
EE	Estonia	LR	Liberia	SG	Singapore		

TITLE

PLANT SUG1 HOMOLOGS

FIELD OF THE INVENTION

5 This invention is in the field of plant molecular biology. More specifically, this invention pertains to nucleic acid fragments encoding coactivator proteins involved in regulation of gene expression in plants and seeds. These same proteins also function as regulatory components of the 26S proteasome which is involved in regulation of protein turnover.

BACKGROUND OF THE INVENTION

10 Gene expression levels are influenced by the interactions of transcription factors with proteins that are present in general transcription complexes. In addition, other non-DNA binding proteins known as coactivators interact with transcription factors and transcription complex proteins to further stimulate transcription. These coactivators are thought to be mediators of the transcription response. One type of coactivator is represented by the SUG1
15 gene, which has been identified in several types of eucaryotic cells such as mouse, yeast and *Xenopus* cells. SUG1 proteins have a coiled coil domain and an "AAA" domain (i.e., ATPases associated with a variety of cellular activities; see vom Bauer et al. (1996) *EMBO J.* 15:110-124).

SUG1 proteins have been shown to bind to the activation domains of the transcription
20 factors, for example yeast SUG1 has been shown to bind to GAL4 of yeast and VP16 of herpesvirus (Swaffield et al. (1995) *Nature* 374:88-91). A mutant GAL4 that lacks the activation domain and therefore cannot activate transcription recovers this function in the presence of a mutant yeast SUG1, indicating a coactivation role for SUG1 (Swaffield et al. (1992) *Nature* 357:698-700). Yeast SUG1 has also been shown to associate with the
25 TATA-binding protein of the TFIID transcription complex *in vivo* and *in vitro*, indicating an intermediary role in gene activation (Swaffield et al. (1995) *Nature* 374:88-91).

Mouse SUG1 has been shown to be able to complement a yeast SUG1 mutant, suggesting that the mouse SUG1 also functions as a transcription coactivator (vom Bauer et al. *supra*). The human thyroid receptor interacting protein 1 (Trip1), which differs from the
30 mouse SUG1 protein by only three amino acids, also complements a yeast SUG1 mutant and binds to the Gal4 and VP16 activation domains (Lee et al. (1995) *Nature* 374:91-94). Interestingly, Trip1 also binds to the thyroid-hormone receptor and the retinoid-X receptor in ligand-dependent fashion which further suggests that Trip1 plays a role in mediating transcription.

35 The mouse SUG1 protein may also play a role in protein degradation. Amino acid comparisons indicate that the mouse SUG1 sequence differs by only a single amino acid residue from the human p45 protein, which was isolated as a subunit of the PA700 proteasome regulatory complex (vom Bauer et al. (1996) *EMBO J* 15:110-124). PA700 includes a set of regulatory proteins that are associated with the 26S proteasome, a complex

that carries out selective degradation of abnormal proteins and naturally short-lived proteins related to cell cycle control and metabolic regulation (Akiyama et al. (1995) *FEBS Letters* 363:151-156).

To date no SUG1 protein homologs have been reported in plants. Because these proteins are involved in transcriptional regulation they are obvious targets for manipulating gene regulation in eukaryotes and as possible targets for screening assays for crop protection chemicals. There is a great deal of interest in identifying the genes that encode SUG1 homologs in plants. These genes may be used to express SUG1 proteins in plant cells to manipulate gene expression. Accordingly, the availability of nucleic acid sequences encoding all or a portion of a SUG1 protein would facilitate studies to better understand and alter gene expression in plants.

SUMMARY OF THE INVENTION

The instant invention relates to an isolated nucleic acid fragment encoding a plant coactivator involved in regulation of gene expression. Moreover, the protein encoded by the isolated nucleic acid fragment also regulates protein turnover. More particularly, this invention concerns isolated nucleic acid fragments encoding plant (specifically from maize, soybean, rice and wheat) homologs of the mouse SUG1 transcriptional coactivator. In addition, this invention relates to a nucleic acid fragment that is complementary to a nucleic acid fragment encoding a plant SUG1 protein.

In another embodiment, the instant invention relates to a chimeric gene that comprises a nucleic acid fragment encoding a plant SUG1 protein, or to a chimeric gene that comprises a nucleic acid fragment that is complementary to a nucleic acid fragment encoding a plant SUG1 protein, the nucleic acid fragment operably linked to suitable regulatory sequences, wherein expression of the chimeric gene results in production of levels of the encoded protein in transformed host cells that are altered (i.e., increased or decreased) relative to the levels produced in untransformed host cells.

In a further embodiment, the instant invention concerns a transformed host cell comprising in its genome a chimeric gene comprising a nucleic acid fragment encoding a plant SUG1 protein or a chimeric gene comprising a nucleic acid fragment that is complementary to the nucleic acid fragment encoding a plant SUG1 protein, the chimeric gene operably linked to suitable regulatory sequences. Expression of the chimeric gene results in production of altered levels of protein encoded by the operably linked nucleic acid fragment in the transformed host cell. The transformed host cell can be of eukaryotic or prokaryotic origin, and include cells derived from higher plants and microorganisms. The invention also includes transformed plants that arise from transformed host cells of higher plants, and seeds derived from such transformed plants.

An additional embodiment of the instant invention concerns a method of altering the level of expression of a plant SUG1 protein in a transformed host cell comprising:

a) transforming a host cell with a chimeric gene comprising a nucleic acid fragment encoding

a plant SUG1 protein or a chimeric gene that comprises a nucleic acid fragment that is complementary to the nucleic acid fragment encoding a plant SUG1 protein; and b) growing the transformed host cell under conditions that are suitable for expression of the chimeric gene wherein expression of the chimeric gene results in production of altered levels of the protein encoded by the operably linked nucleic acid fragment in the transformed host cell.

An addition embodiment of the instant invention concerns a method for obtaining a nucleic acid fragment encoding all or substantially all of an amino acid sequence encoding a plant SUG1 protein.

A further embodiment of the instant invention is a method for evaluating at least one compound for its ability to inhibit the activity of a plant SUG1 protein, the method comprising the steps of: (a) transforming a host cell with a chimeric gene comprising a nucleic acid fragment encoding a plant SUG1 protein operably linked to suitable regulatory sequences; (b) growing the transformed host cell under conditions that are suitable for expression of the chimeric gene wherein expression of the chimeric gene results in production of the plant SUG1 protein in the transformed host cell; (c) optionally purifying the plant SUG1 protein expressed by the transformed host cell; (d) treating the plant SUG1 protein with a compound to be tested; and (e) comparing the activity of the plant SUG1 protein that has been treated with a test compound to the activity of an untreated plant SUG1 protein, thereby selecting compounds with potential for inhibitory activity.

BRIEF DESCRIPTION OF THE DRAWINGS AND SEQUENCE DESCRIPTIONS

The invention can be more fully understood from the following detailed description and the accompanying drawings and sequence descriptions which form a part of this application.

Figure 1 shows a comparison of the amino acid sequences of the mouse SUG1 protein (Z54219), the human p45 protein (P47210), the human Tripl transcriptional coactivator (L38810), the *Saccharomyces cerevisiae* SUG1 transcriptional coactivator (X66400), and the instant soybean and maize SUG1 protein homologs (se1.pk0023.b5 and cs1.pk0051.b7, respectively).

Figure 2 shows a comparison of the amino acid sequences of the mouse SUG1 protein SEQ ID NO:7 (3), the instant soybean clone se1.pk0023.b5 (2), a corn contig (cs1.pk0051.b7, cr1n.pk0096.b2 and ct1n.pk0056.c11) (1) and a wheat clone (wl1n.pk0053.g3) (4).

The following sequence descriptions and sequence listings attached hereto comply with the rules governing nucleotide and/or amino acid sequence disclosures in patent applications as set forth in 37 C.F.R. §1.821-1.825.

SEQ ID NO:1 is the nucleotide sequence comprising the entire cDNA insert in clone se1.pk0023.b5 encoding a soybean SUG1 protein.

SEQ ID NO:2 is the deduced amino acid sequence of a soybean SUG1 homolog derived from the nucleotide sequence of SEQ ID NO:1.

SEQ ID NO:3 is the nucleotide sequence comprising the entire cDNA insert in clone cs1.pk0051.b7 encoding corn SUG1 protein.

5 SEQ ID NO:4 is the deduced amino acid sequence of a portion of a maize SUG1 homolog derived from the nucleotide sequence of SEQ ID NO:3.

SEQ ID NO:5 is the nucleotide sequence comprising a contig assembled from the cDNA inserts in clones cs1.pk0051.b7, cr1n.pk0096.b2 and ctaln.pk0056.c11 encoding a corn SUG1 protein.

10 SEQ ID NO:6 is the deduced amino acid sequence of a SUG1 protein derived from the nucleotide sequence of SEQ ID NO:5.

SEQ ID NO:7 is the amino acid sequence encoding the mouse SUG1 protein having EMBL accession No. Z54219.

15 SEQ ID NO:8 is the amino acid sequence encoding the *Saccharomyces cerevisiae* SUG1 transcriptional coactivator having EMBL accession No. X66400.

SEQ ID NO:9 is the amino acid sequence encoding the human Tripl transcriptional coactivator having GenBank accession No. L38810.

SEQ ID NO:10 is the amino acid sequence encoding the human p45 protein having SWISS-PROT accession No. P47210.

20 SEQ ID NO:11 is the nucleotide sequence comprising a portion of a cDNA insert in clone wl1n.pk0053.g3 encoding a wheat SUG1.

SEQ ID NO:12 is the deduced amino acid sequence of a wheat SUG1 protein from the nucleotide sequence of SEQ ID NO:11.

25 SEQ ID NO:13 is the nucleotide sequence comprising a portion of a cDNA insert in clone rlr6.pk0064.e10 encoding a rice 26S protease regulatory subunit 8.

SEQ ID NO:14 is the deduced amino acid sequence of a 26S protease regulatory subunit 8 protein from the nucleotide sequence of SEQ ID NO:13.

30 The Sequence Descriptions contain the one letter code for nucleotide sequence characters and the three letter codes for amino acids as defined in conformity with the IUPAC-IYUB standards described in *Nucleic Acids Research* 13:3021-3030 (1985) and in the *Biochemical Journal* 219 (No. 2):345-373 (1984) which are herein incorporated by reference. The symbols and format used for nucleotide and amino acid sequence data comply with the rules set forth in 37 C.F.R. §1.822.

DETAILED DESCRIPTION OF THE INVENTION

35 The amino acid sequence similarity between the instant plant SUG1 proteins and the mouse and yeast SUG1 proteins, human Tripl and p45 proteins indicates that the plant SUG1 proteins may function as transcriptional coactivators and as regulators of protein degradation (Figure 1). The plant proteins may therefore be used to reduce expression of specific genes whose promoters are normally regulated by SUG1, using antisense or

co-suppression technology. The instant plant proteins may also be used to enhance gene expression of those genes whose promoters are normally targeted by the transcription factors that the plant SUG1 proteins normally interact with.

Alternatively, the plant SUG1 protein coactivation function can be targeted to a novel promoter region by the addition of either a DNA binding domain or a protein-protein interaction domain. The instant plant SUG1 protein can be fused to a very defined DNA-binding domain, such as, but not limited to, a bacterial *lexA* DNA binding domain, a yeast Gal4 DNA-binding domain or a DNA binding domain from a plant transcription factor. On the other hand, a synthetic promoter can be designed to contain multiple copies of a target site which is necessary for the specific binding by either the *lexA*, Gal4 or plant DNA binding domain. By using this approach, the plant SUG1 protein can be specifically targeted to the engineered synthetic promoter, thus leading to a higher level of gene expression when used in combination with an interacting transcription factor.

Additionally, plant SUG1 proteins can be fused to a transcription factor that already includes its own DNA binding domain in order to target the coactivator. Besides DNA-binding domains, the plant SUG1 proteins can also be fused to other transcription regulatory proteins, such as transcription mediators. Normally, these mediators do not bind to DNA directly and are recruited to their target sites by interaction with other DNA-binding proteins. By fusing the plant SUG1 protein to these mediators, the plant SUG1 protein can be targeted to specific regulatory elements through the interaction between the mediators and other DNA-binding proteins.

In the context of this disclosure, a number of terms shall be utilized. As used herein, an "isolated nucleic acid fragment" is a polymer of RNA or DNA that is single- or double-stranded, optionally containing synthetic, non-natural or altered nucleotide bases. An isolated nucleic acid fragment in the form of a polymer of DNA may be comprised of one or more segments of cDNA, genomic DNA or synthetic DNA. As used herein, "contig" refers to an assemblage of overlapping nucleic acid sequences to form one contiguous nucleotide sequence. For example, several DNA sequences can be compared and aligned to identify common or overlapping regions. The individual sequences can then be assembled into a single contiguous nucleotide sequence.

As used herein, "substantially similar" refers to nucleic acid fragments wherein changes in one or more nucleotide bases results in substitution of one or more amino acids, but do not affect the functional properties of the protein encoded by the DNA sequence. "Substantially similar" also refers to nucleic acid fragments wherein changes in one or more nucleotide bases does not affect the ability of the nucleic acid fragment to mediate alteration of gene expression by antisense or co-suppression technology. "Substantially similar" also refers to modifications of the nucleic acid fragments of the instant invention such as deletion or insertion of one or more nucleotides that do not substantially affect the functional properties of the resulting transcript vis-à-vis the ability to mediate alteration of gene

expression by antisense or co-suppression technology or alteration of the functional properties of the resulting protein molecule. It is therefore understood that the invention encompasses more than the specific exemplary sequences.

For example, it is well known in the art that antisense suppression and co-suppression
5 of gene expression may be accomplished using nucleic acid fragments representing less than the entire coding region of a gene, and by nucleic acid fragments that do not share 100% sequence identity with the gene to be suppressed. Moreover, alterations in a gene which result in the production of a chemically equivalent amino acid at a given site, but do not effect the functional properties of the encoded protein, are well known in the art. Thus, a
10 codon for the amino acid alanine, a hydrophobic amino acid, may be substituted by a codon encoding another less hydrophobic residue, such as glycine, or a more hydrophobic residue, such as valine, leucine, or isoleucine. Similarly, changes which result in substitution of one negatively charged residue for another, such as aspartic acid for glutamic acid, or one
15 positively charged residue for another, such as lysine for arginine, can also be expected to produce a functionally equivalent product. Nucleotide changes which result in alteration of the N-terminal and C-terminal portions of the protein molecule would also not be expected to alter the activity of the protein. Each of the proposed modifications is well within the routine skill in the art, as is determination of retention of biological activity of the encoded products. Moreover, the skilled artisan recognizes that substantially similar nucleic acid
20 sequences encompassed by this invention are also defined by their ability to hybridize, under stringent conditions (0.1X SSC, 0.1% SDS, 65°C), with the sequences exemplified herein. Preferred substantially similar nucleic acid fragments of the instant invention are those nucleic acid fragments whose DNA sequences are 85% identical to the coding sequence of the nucleic acid fragments reported herein. More preferred nucleic acid fragments are 90%
25 identical to the coding sequence of the nucleic acid fragments reported herein. Most preferred are nucleic acid fragments that are 95% identical to the coding sequence of the nucleic acid fragments reported herein. The percent identity used herein, can be precisely determined by the DNASTAR protein alignment protocol using the Hein algorithm (Hein, J.J. (1990) Unified Approach to Alignment and Phylogenies. *Methods in Enzymology*,
30 vol. 183, 626-645). Default parameters for the J. J. Hein method for multiple alignments are: GAP PENALTY=11, GAP LENGTH PENALTY=3; for pairwise alignments KTUPLE 6.

A "substantial portion" of an amino acid or nucleotide sequence comprises enough of the amino acid sequence of a polypeptide or the nucleotide sequence of a gene to afford putative identification of that polypeptide or gene, either by manual evaluation of the
35 sequence by one skilled in the art, or by computer-automated sequence comparison and identification using algorithms such as BLAST (Basic Local Alignment Search Tool; Altschul, S. F., et al., (1993) *J. Mol. Biol.* 215:403-410; see also www.ncbi.nlm.nih.gov/BLAST/). In general, a sequence of ten or more contiguous amino acids or thirty or more nucleotides is necessary in order to putatively identify a polypeptide

or nucleic acid sequence as homologous to a known protein or gene. Moreover, with respect to nucleotide sequences, gene specific oligonucleotide probes comprising 20-30 contiguous nucleotides may be used in sequence-dependent methods of gene identification (e.g., Southern hybridization) and isolation (e.g., *in situ* hybridization of bacterial colonies or bacteriophage plaques). In addition, short oligonucleotides of 12-15 bases may be used as amplification primers in PCR in order to obtain a particular nucleic acid fragment comprising the primers. Accordingly, a "substantial portion" of a nucleotide sequence comprises enough of the sequence to afford specific identification and/or isolation of a nucleic acid fragment comprising the sequence. The instant specification teaches partial or complete amino acid and nucleotide sequences encoding a particular plant protein. The skilled artisan, having the benefit of the sequences as reported herein, may now use all or a substantial portion of the disclosed sequences for purposes known to those skilled in this art. Accordingly, the instant invention comprises the complete sequences as reported in the accompanying Sequence Listing, as well as substantial portions of those sequences as defined above.

"Codon degeneracy" refers to divergence in the genetic code permitting variation of the nucleotide sequence without effecting the amino acid sequence of an encoded polypeptide. Accordingly, the instant invention relates to any nucleic acid fragment that encodes all or a substantial portion of the amino acid sequence encoding a corn, soybean, rice or wheat SUG1 protein as set forth in SEQ ID NOs:2, 4, 6, 12 and 14. The skilled artisan is well aware of the "codon-bias" exhibited by a specific host cell in usage of nucleotide codons to specify a given amino acid. Therefore, when synthesizing a gene for improved expression in a host cell, it is desirable to design the gene such that its frequency of codon usage approaches the frequency of preferred codon usage of the host cell.

"Synthetic genes" can be assembled from oligonucleotide building blocks that are chemically synthesized using procedures known to those skilled in the art. These building blocks are ligated and annealed to form gene segments which are then enzymatically assembled to construct the entire gene. "Chemically synthesized", as related to a sequence of DNA, means that the component nucleotides were assembled *in vitro*. Manual chemical synthesis of DNA may be accomplished using well established procedures, or automated chemical synthesis can be performed using one of a number of commercially available machines. Accordingly, the genes can be tailored for optimal gene expression based on optimization of nucleotide sequence to reflect the codon bias of the host cell. The skilled artisan appreciates the likelihood of successful gene expression if codon usage is biased towards those codons favored by the host. Determination of preferred codons can be based on a survey of genes derived from the host cell where sequence information is available.

"Gene" refers to a nucleic acid fragment that encodes a specific protein, including regulatory sequences preceding (5' non-coding sequences) and following (3' non-coding sequences) the coding sequence. "Native gene" refers to a gene as found in nature with its

own regulatory sequences. "Chimeric gene" refers any gene that is not a native gene, comprising regulatory and coding sequences that are not found together in nature.

Accordingly, a chimeric gene may comprise regulatory sequences and coding sequences that are derived from different sources, or regulatory sequences and coding sequences derived
5 from the same source, but arranged in a manner different than that found in nature.

"Endogenous gene" refers to a native gene in its natural location in the genome of an organism. A "foreign" gene refers to a gene not normally found in the host organism, but that is introduced into the host organism by gene transfer. Foreign genes can comprise native genes inserted into a non-native organism, or chimeric genes. A "transgene" is a gene
10 that has been introduced into the genome by a transformation procedure.

"Coding sequence" refers to a DNA sequence that codes for a specific amino acid sequence. "Regulatory sequences" refer to nucleotide sequences located upstream (5' non-coding sequences), within, or downstream (3' non-coding sequences) of a coding sequence, and which influence the transcription, RNA processing or stability, or translation of the
15 associated coding sequence. Regulatory sequences may include promoters, translation leader sequences, introns, and polyadenylation recognition sequences.

"Promoter" refers to a DNA sequence capable of controlling the expression of a coding sequence or functional RNA. In general, a coding sequence is located 3' to a promoter sequence. The promoter sequence consists of proximal and more distal upstream
20 elements, the latter elements often referred to as enhancers. Accordingly, an "enhancer" is a DNA sequence which can stimulate promoter activity and may be an innate element of the promoter or a heterologous element inserted to enhance the level or tissue-specificity of a promoter. Promoters may be derived in their entirety from a native gene, or be composed of different elements derived from different promoters found in nature, or even comprise
25 synthetic DNA segments. It is understood by those skilled in the art that different promoters may direct the expression of a gene in different tissues or cell types, or at different stages of development, or in response to different environmental conditions. Promoters which cause a gene to be expressed in most cell types at most times are commonly referred to as "constitutive promoters". New promoters of various types useful in plant cells are
30 constantly being discovered; numerous examples may be found in the compilation by Okamuro and Goldberg, (1989) *Biochemistry of Plants* 15:1-82. It is further recognized that since in most cases the exact boundaries of regulatory sequences have not been completely defined, DNA fragments of different lengths may have identical promoter activity.

The "translation leader sequence" refers to a DNA sequence located between the
35 promoter sequence of a gene and the coding sequence. The translation leader sequence is present in the fully processed mRNA upstream of the translation start sequence. The translation leader sequence may affect processing of the primary transcript to mRNA, mRNA stability or translation efficiency. Examples of translation leader sequences have been described (Turner, R. and Foster, G.D. (1995) *Molecular Biotechnology* 3:225).

The "3' non-coding sequences" refer to DNA sequences located downstream of a coding sequence and include polyadenylation recognition sequences and other sequences encoding regulatory signals capable of affecting mRNA processing or gene expression. The polyadenylation signal is usually characterized by affecting the addition of polyadenylic acid tracts to the 3' end of the mRNA precursor. The use of different 3' non-coding sequences is exemplified by Ingelbrecht et al., (1989) *Plant Cell* 1:671-680.

"RNA transcript" refers to the product resulting from RNA polymerase-catalyzed transcription of a DNA sequence. When the RNA transcript is a perfect complementary copy of the DNA sequence, it is referred to as the primary transcript. Alternatively, the RNA transcript may be an RNA sequence derived from posttranscriptional processing of the primary transcript; this is referred to as the mature RNA. "Messenger RNA" (mRNA) refers to the RNA that is without introns and that can be translated into protein by the cell. "cDNA" refers to a double-stranded DNA that is complementary to and derived from mRNA. "Sense" RNA refers to an RNA transcript that includes the mRNA and so can be translated into protein by the cell. "Antisense RNA" refers to a RNA transcript that is complementary to all or part of a target primary transcript or mRNA and that blocks the expression of a target gene (U.S. Pat. No. 5,107,065). The complementarity of an antisense RNA may be with any part of the specific gene transcript, i.e., at the 5' non-coding sequence, 3' non-coding sequence, introns, or the coding sequence. "Functional RNA" refers to antisense RNA, ribozyme RNA, or other RNA that is not translated yet has an effect on cellular processes.

The term "operably linked" refers to the association of nucleic acid sequences on a single nucleic acid fragment so that the function of one is affected by the other. For example, a promoter is operably linked with a coding sequence when it is capable of affecting the expression of that coding sequence (i.e., that the coding sequence is under the transcriptional control of the promoter). Coding sequences can be operably linked to regulatory sequences in sense or antisense orientation.

The term "expression", as used herein, refers to the transcription and stable accumulation of sense (mRNA) or antisense RNA derived from the nucleic acid fragment of the invention. Expression may also refer to translation of mRNA into a polypeptide. "Antisense inhibition" refers to the production of antisense RNA transcripts capable of suppressing the expression of the target protein. "Overexpression" refers to the production of a gene product in transgenic organisms that exceeds levels of production in normal or non-transformed organisms. "Co-suppression" refers to the production of sense RNA transcripts capable of suppressing the expression of identical or substantially similar foreign or endogenous genes (U.S. Pat. No. 5,231,020).

"Altered levels" refers to the production of gene product(s) in transgenic organisms in amounts or proportions that differ from that of normal or non-transformed organisms.

"Transformation" refers to the transfer of a nucleic acid fragment into the genome of a host organism, resulting in genetically stable inheritance. Host organisms containing the transformed nucleic acid fragments are referred to as "transgenic" organisms. Examples of methods of plant transformation include Agrobacterium-mediated transformation (De Blaere et al. (1987) *Meth. Enzymol.* 143:277) and particle-accelerated or "gene gun" transformation technology (Klein et al. (1987) *Nature (London)* 327:70-73; U.S. Pat. No. 4,945,050).

Standard recombinant DNA and molecular cloning techniques used herein are well known in the art and are described more fully in Sambrook, J., Fritsch, E.F. and Maniatis, T. *Molecular Cloning: A Laboratory Manual*; Cold Spring Harbor Laboratory Press: Cold Spring Harbor, 1989 (hereinafter "Maniatis").

This invention relates to plant cDNAs with homology to the mouse, yeast and *Xenopus* SUG1 proteins. The cDNAs have been isolated and identified by comparison of random plant cDNA sequences to several publically available databases using the BLAST algorithms well known to those skilled in the art. The nucleotide sequence of a soybean cDNA clone (se1.pk0023.b5) encoding a SUG1 homolog is provided in SEQ ID NO:1, and the deduced amino acid sequence is provided in SEQ ID NO:2. The nucleotide sequence of a maize SUG1 homolog (cs1.pk0051.b7) is provided in SEQ ID NOs:3 and 5, and the deduced amino acid sequence is provided in SEQ ID NO:4 and 6. The nucleotide sequence of a wheat SUG1 homolog (wl1n.pk0053.g3) is provided in SEQ ID NO:11, and the deduced amino acid sequence is provided in SEQ ID NO:12. Lastly, the nucleotide sequence of a rice 26S protease regulatory subunit 8 homolog (rlr6.pk0064.e10) is provided in SEQ ID NO:13, and the deduced amino acid sequence is provided in SEQ ID NO:14. Homologs of these proteins from other plants can now be identified by comparison of random cDNA sequences to the maize, soybean, and wheat sequences provided herein.

The full insert of cDNA clone se1.pk0023.b5 encoding the soybean SUG1 homolog has been completely sequenced. Amino acid sequence comparison indicates that there is 80% sequence identity between this soybean homolog and the mouse SUG1 protein (Figure 1). Sequence alignments and percent identity calculations were performed by the Jotun Hein method using the Megalign program of DNASTar™ sequence analysis software (DNASTAR Inc. 1228 South Park Street, Madison Wisconsin, 53715)

Based on the high sequence homology, it is believed that this soybean clone encodes a plant homolog of mouse SUG1. This is the first indication that there is a similar SUG-mediated gene activation system in plants.

Likewise, the full insert of cDNA clone cs1.pk0051.b7 encoding the maize SUG1 homolog has been completely sequenced. The region encoding approximately thirty amino acids is missing from the 5' end of the cDNA insert. At the amino acid level, this maize peptide is approximately 95% identical to the soybean SUG1 protein encoded by cDNA clone se1.pk0023.b5. Nucleotide identity between the soybean and maize cDNAs is approximately 78%. Sequence alignments and percent identity calculations were performed

by the Jotun Hein method using the Megalign program of DNASTar™ sequence analysis software (DNASTAR Inc. 1228 South Park Street, Madison Wisconsin, 53715).

The nucleic acid fragments of the instant invention may be used to isolate cDNAs and genes encoding other homologs of SUG1 from the same or other plant species. Isolation of homologous genes using sequence-dependent protocols is well known in the art. Examples of sequence-dependent protocols include, but are not limited to, methods of nucleic acid hybridization, and methods of DNA and RNA amplification as exemplified by various uses of nucleic acid amplification technologies (e.g., polymerase chain reaction, ligase chain reaction).

For example, genes encoding other plant homologs of SUG1 either as cDNAs or genomic DNAs, could be isolated directly by using all or a portion of the instant nucleic acid fragments as DNA hybridization probes to screen libraries from any desired plant employing methodology well known to those skilled in the art. Specific oligonucleotide probes based upon the instant nucleic acid sequences can be designed and synthesized by methods known in the art (Maniatis). Moreover, the entire sequences can be used directly to synthesize DNA probes by methods known to the skilled artisan such as random primers DNA labeling, nick translation, or end-labeling techniques, or RNA probes using available *in vitro* transcription systems. In addition, specific primers can be designed and used to amplify a part of or full-length of the instant sequences. The resulting amplification products can be labeled directly during amplification reactions or labeled after amplification reactions, and used as probes to isolate full length cDNA or genomic fragments under conditions of appropriate stringency. Genomic fragments can be isolated that include the promoter region that directs expression of the plant SUG1 protein. This promoter may be prepared as a DNA fragment including regulatory elements with or without the untranslated leader and used in expression of other coding regions or for co-suppression.

In addition, two short segments of the instant nucleic acid fragments may be used in polymerase chain reaction protocols to amplify longer nucleic acid fragments encoding homologous genes from DNA or RNA. The polymerase chain reaction may also be performed on a library of cloned nucleic acid fragments wherein the sequence of one primer is derived from the instant nucleic acid fragments, and the sequence of the other primer takes advantage of the presence of the polyadenylic acid tracts to the 3' end of the mRNA precursor encoding plant genes. Alternatively, the second primer sequence may be based upon sequences derived from the cloning vector. For example, the skilled artisan can follow the RACE protocol (Frohman et al., (1988) *PNAS USA* 85:8998) to generate cDNAs by using PCR to amplify copies of the region between a single point in the transcript and the 3' or 5' end. Primers oriented in the 3' and 5' directions can be designed from the instant sequences. Using commercially available 3' RACE or 5' RACE systems (BRL), specific 3' or 5' cDNA fragments can be isolated (Ohara et al., (1989) *PNAS USA* 86:5673; Loh et al., (1989) *Science* 243:217). Products generated by the 3' and 5' RACE procedures can be

combined to generate full-length cDNAs (Frohman, M.A. and Martin, G.R., (1989) *Techniques* 1:165).

Availability of the instant nucleotide and deduced amino acid sequences facilitates immunological screening of cDNA expression libraries. Synthetic peptides representing portions of the instant amino acid sequences may be synthesized. These peptides can be used to immunize animals to produce polyclonal or monoclonal antibodies with specificity for peptides or proteins comprising the amino acid sequences. These antibodies can be then be used to screen cDNA expression libraries to isolate full-length cDNA clones of interest (Lerner, R.A. (1984) *Adv. Immunol.* 36:1; Maniatis).

The nucleic acid fragments of the instant invention may be used to create transgenic plants in which the disclosed plant homologs of SUG1 are present at higher or lower levels than normal or in cell types or developmental stages in which it is not normally found. This would have the effect of altering the level of a plant SUG1 protein in those cells.

Overexpression of plant a SUG1 protein may be accomplished by first constructing a chimeric gene in which the coding region is operably linked to a promoter capable of directing expression of a gene in the desired tissues at the desired stage of development. For reasons of convenience, the chimeric gene may comprise a promoter sequence and translation leader sequence derived from the same gene. A 3' non-coding sequence encoding a transcription termination signal may also be provided. The instant chimeric gene may also comprise one or more introns in order to facilitate gene expression.

Plasmid vectors comprising the instant chimeric gene can then constructed. The choice of plasmid vector is dependent upon the method that will be used to transform host plants. The skilled artisan is well aware of the genetic elements that must be present on the plasmid vector in order to successfully transform, select and propagate host cells containing the chimeric gene. The skilled artisan will also recognize that different independent transformation events will result in different levels and patterns of expression (Jones et al., (1985) *EMBO J.* 4:2411-2418; De Almeida et al., (1989) *Mol. Gen. Genetics* 218:78-86), and thus that multiple events must be screened in order to obtain lines displaying the desired expression level and pattern. Such screening may be accomplished by Southern analysis of DNA, Northern analysis of mRNA expression, Western analysis of protein expression, or phenotypic analysis.

For some applications, it may be desirable to reduce or eliminate expression of the genes encoding the instant plant of SUG1 homologs. In order to accomplish this, chimeric genes designed for co-suppression of the instant plant SUG1 protein genes can be constructed by linking the genes or gene fragments encoding the plant SUG1 proteins to plant promoter sequences. Alternatively, chimeric genes designed to express antisense RNA for all or part of the instant nucleic acid fragments can be constructed by linking the genes or gene fragments in reverse orientation to plant promoter sequences. Either the co-suppression or antisense chimeric genes could be introduced into plants via

transformation wherein expression of the corresponding endogenous genes are reduced or eliminated.

5 The instant plant SUG1 homologs (or portions thereof) may be produced in heterologous host cells, particularly in the cells of microbial hosts, and can be used to prepare antibodies to the SUG1 proteins by methods well known to those skilled in the art. The antibodies are useful for detecting plant SUG1 proteins *in situ* in cells or *in vitro* in cell extracts. Preferred heterologous host cells for production of the instant SUG1 proteins are microbial hosts. Microbial expression systems and expression vectors containing regulatory sequences that direct high level expression of foreign proteins are well known to those skilled in the art. Any of these could be used to construct chimeric genes for production of the instant plant SUG1 homologs. These chimeric genes could then be introduced into appropriate microorganisms via transformation to provide high level expression of corn, soybean or wheat SUG1 proteins. An example of a vector for high level expression of the instant plant SUG1 homologs in a bacterial host is provided (Example 6).

15 Additionally, the instant SUG1 proteins can be used as targets to facilitate design and/or identification of inhibitors of the activity of the protein that may be useful as herbicides. This is desirable because the protein described plays a key role in regulation of gene expression. Accordingly, inhibition of the activity of the proteins described herein could lead to inhibition of gene expression sufficient to inhibit plant growth. Thus, the instant SUG1 proteins could be appropriate for new herbicide discovery and design.

20 All or a portion of the nucleic acid fragments of the instant invention may also be used as probes for genetically and physically mapping the genes that they are a part of, and as markers for traits linked to expression of the instant plant SUG1 proteins. Such information may be useful in plant breeding in order to develop lines with desired phenotypes.

25 For example, the instant nucleic acid fragments may be used as restriction fragment length polymorphism (RFLP) markers. Southern blots (Maniatis) of restriction-digested plant genomic DNA may be probed with the nucleic acid fragments of the instant invention. The resulting banding patterns may then be subjected to genetic analyses using computer programs such as MapMaker (Lander et al., (1987) *Genomics* 1:174-181) in order to construct a genetic map. In addition, the nucleic acid fragments of the instant invention may be used to probe Southern blots containing restriction endonuclease-treated genomic DNAs of a set of individuals representing parent and progeny of a defined genetic cross. Segregation of the DNA polymorphisms is noted and used to calculate the position of the instant nucleic acid sequence in the genetic map previously obtained using this population

30 (Botstein, D. et al., (1980) *Am.J.Hum.Genet.* 32:314-331).

35 The production and use of plant gene-derived probes for use in genetic mapping is described in R. Bernatzky, R. and Tanksley, S.D. (1986) *Plant Mol.Biol.Reporter* 4(1):37-41. Numerous publications describe genetic mapping of specific cDNA clones using the methodology outlined above or variations thereof. For example, F2 intercross

populations, backcross populations, randomly mated populations, near isogenic lines, and other sets of individuals may be used for mapping. Such methodologies are well known to those skilled in the art.

5 Nucleic acid probes derived from the instant nucleic acid sequences may also be used for physical mapping (i.e., placement of sequences on physical maps; see Hoheisel, J. D., et al., In: *Nonmammalian Genomic Analysis: A Practical Guide*, Academic press 1996, pp. 319-346, and references cited therein).

10 In another embodiment, nucleic acid probes derived from the instant nucleic acid sequences may be used in direct fluorescence *in situ* hybridization (FISH) mapping (Trask, B. J. (1991) *Trends Genet.* 7:149-154). Although current methods of FISH mapping favor use of large clones (several to several hundred KB; see Laan, M. et al. (1995) *Genome Research* 5:13-20), improvements in sensitivity may allow performance of FISH mapping using shorter probes.

15 A variety of nucleic acid amplification-based methods of genetic and physical mapping may be carried out using the instant nucleic acid sequences. Examples include allele-specific amplification (Kazazian, H. H. (1989) *J. Lab. Clin. Med.* 114(2):95-96), polymorphism of PCR-amplified fragments (CAPS; Sheffield, V. C. et al. (1993) *Genomics* 16:325-332), allele-specific ligation (Landegren, U. et al. (1988) *Science* 241:1077-1080), nucleotide extension reactions (Sokolov, B. P. (1990) *Nucleic Acid Res.* 18:3671), Radiation
20 Hybrid Mapping (Walter, M. A. et al. (1997) *Nature Genetics* 7:22-28) and Happy Mapping (Dear, P. H. and Cook, P. R. (1989) *Nucleic Acid Res.* 17:6795-6807). For these methods, the sequence of a nucleic acid fragment is used to design and produce primer pairs for use in the amplification reaction or in primer extension reactions. The design of such primers is well known to those skilled in the art. In methods employing PCR-based genetic mapping,
25 it may be necessary to identify DNA sequence differences between the parents of the mapping cross in the region corresponding to the instant nucleic acid sequence. This, however, is generally not necessary for mapping methods.

30 Loss of function mutant phenotypes may be identified for the instant cDNA clones either by targeted gene disruption protocols or by identifying specific mutants for these genes contained in a maize population carrying mutations in all possible genes (Ballinger and Benzer, (1989) *Proc. Natl. Acad. Sci USA* 86:9402; Koes et al., (1995) *Proc. Natl. Acad. Sci USA* 92:8149; Bensen et al., (1995) *Plant Cell* 7:75). The latter approach may be accomplished in two ways. First, short segments of the instant nucleic acid fragments may be used in polymerase chain reaction protocols in conjunction with a mutation tag sequence
35 primer on DNAs prepared from a population of plants in which Mutator transposons or some other mutation-causing DNA element has been introduced (see Bensen, *supra*). The amplification of a specific DNA fragment with these primers indicates the insertion of the mutation tag element in or near the plant SUG1 gene. Alternatively, the SUG1 gene may be used as a hybridization probe against PCR amplification products generated from the

mutation population using the mutation tag sequence primer in conjunction with an arbitrary genomic site primer, such as that for a restriction enzyme site-anchored synthetic adaptor. With either method, a plant containing a mutation in the endogenous SUG1 gene can be identified and obtained. This mutant plant can then be used to determine or confirm the natural function of the plant SUG1 gene product.

EXAMPLES

The present invention is further defined in the following Examples, in which all parts and percentages are by weight and degrees are Celsius, unless otherwise stated. It should be understood that these Examples, while indicating preferred embodiments of the invention, are given by way of illustration only. From the above discussion and these Examples, one skilled in the art can ascertain the essential characteristics of this invention, and without departing from the spirit and scope thereof, can make various changes and modifications of the invention to adapt it to various usages and conditions.

EXAMPLE 1

Composition of cDNA Libraries; Isolation and Sequencing of cDNA Clones

cDNA libraries representing mRNAs from soybean embryo obtained seven days after flowering and maize leaf sheath obtained from five week old *Zea mays* B73 maize plants were prepared. cDNA libraries representing mRNAs from wheat leaf from 7 day old etiolated seedlings (normalized, normalized essentially as described in U.S. Pat. No. 5,482,845) were prepared. A cDNA library representing mRNAs from rice leaf 15 days after germination and 6 hours after infection of strain *Magaporthe grisea* 4360-R-62 (AVR2-YAMO) was also prepared.

Separate cDNA libraries representing the maize, soybean, rice and wheat cDNAs were prepared in Uni-ZAP™ XR vectors according to the manufacturer's protocol (Stratagene Cloning Systems, La Jolla, CA). Conversion of the Uni-ZAP™ XR libraries into plasmid libraries was accomplished according to the protocol provided by Stratagene. Upon conversion, cDNA inserts were contained in the plasmid vector pBluescript. cDNA inserts from randomly picked bacterial colonies containing recombinant pBluescript plasmids were amplified via polymerase chain reaction using primers specific for vector sequences flanking the inserted cDNA sequences. Amplified insert DNAs were sequenced in dye-primer sequencing reactions to generate partial cDNA sequences (expressed sequence tags or "ESTs"; see Adams, M. D. et al., (1991) *Science* 252:1651). The resulting ESTs were analyzed using a Perkin Elmer Model 377 fluorescent sequencer.

EXAMPLE 2

Identification and Characterization of cDNA Clones

ESTs encoding homologs of the mouse, yeast and *Xenopus* SUG1 proteins were identified by conducting BLAST (Basic Local Alignment Search Tool; Altschul, S. F., et al., (1993) *J. Mol. Biol.* 215:403-410; see also www.ncbi.nlm.nih.gov/BLAST/) searches for similarity to sequences contained in the BLAST "nr" database (comprising all non-

redundant GenBank CDS translations, sequences derived from the 3-dimensional structure Brookhaven Protein Data Bank, the last major release of the SWISS-PROT protein sequence database, EMBL, and DDBJ databases). The cDNA sequences obtained in Example 1 were analyzed for similarity to all publicly available DNA sequences contained in the "nr" database using the BLASTN algorithm provided by the National Center for Biotechnology Information (NCBI). The DNA sequences were translated in all reading frames and compared for similarity to all publicly available protein sequences contained in the "nr" database using the BLASTX algorithm (Gish, W. and States, D. J. (1993) *Nature Genetics* 3:266-272) provided by the NCBI. For convenience, the P-value (probability) of observing a match of a cDNA sequence to a sequence contained in the searched databases merely by chance as calculated by BLAST are reported herein as "pLog" values, which represent the negative of the logarithm of the reported P-value. Accordingly, the greater the pLog value, the greater the likelihood that the cDNA sequence and the BLAST "hit" represent homologous proteins.

EXAMPLE 3

Characterization of cDNA Clones Encoding SUG1 Homologs

The BLASTX search using the EST sequences from clones se1.pk0023.b5, cs1.pk0051.b7, cr1n.pk0096.b2 and ct1n.pk0056.c11 revealed similarity of the proteins encoded by the cDNAs to the SUG1 protein from mouse (EMBL Accession No. Z54219). The BLASTX search using the EST sequence from clone wl1n.pk0053.g3 revealed similarity of the protein encoded by the cDNA to the SUG1 protein from *Xenopus laevis* (GenBank Accession No. X81986). The BLASTX search using the EST sequence from clone rlr6.pk0064.e10 revealed similarity of the protein encoded by the cDNA to the 26S Protease Regulatory Subunit 8 (a SUG1 protein homolog) from *Dictyostelium discoideum* (SWISS PROT Accession No. P34124). The BLAST results for each of these ESTs are shown in Table 3:

TABLE 1
BLAST Results for Clones Encoding Polypeptides Homologous to SUG1 Proteins

Clone	BLAST pLog Score (Organism Accession Number)
se1.pk0023.b5	35.05 (Z54219)
cs1.pk0051.b7	27.85 (Z54219)
Contig composed of:	217.03 (Z54219)
cs1.pk0051.b7	
ta1n.pk0056.c11	
cr1n.pk0096.b2	
wl1n.pk0053.g3	21.62 (X81986)
rlr6.pk0064.e10	53.19 (P34124)

In the process of comparing the corn ESTs it was found that clones cs1.pk0051.b7, cr1n.pk0096.b2 and ctaln.pk0056.c11 had overlapping regions of homology. Using this homology it was possible to align the entire cDNA insert of clone cs1.pk0051.b7, with the ESTs of clones cr1n.pk0096.b2 and ctaln.pk0056.c11 and assemble a contig (a contig is an assemblage of overlapping nucleic acid sequences to form one contiguous nucleotide sequence). The individual sequences were assembled into a unique contiguous nucleotide sequence encoding a unique corn SUG1 protein. The nucleotide sequence of the contig is shown in SEQ ID NO:5; the deduced amino acid sequence of the contig is shown in SEQ ID NO:6. The nucleotide sequence of the cDNA clone w11n.pk0053.g3 is shown in SEQ ID NO:11; the deduced amino acid sequence encoded by clone w11n.pk0053.g3 is shown in SEQ ID NO:12. The nucleotide sequence of the cDNA clone rlr6.pk0064.e10 is shown in SEQ ID NO:13; the deduced amino acid sequence encoded by clone rlr6.pk0064.e10 is shown in SEQ ID NO:14. BLAST scores and probabilities indicate that the instant nucleic acid fragments encode portions of a plant SUG1 protein. These sequences represent plant sequences encoding a SUG1 protein.

The sequence of the entire cDNA insert in clone se1.pk0023.b5 was determined and is shown in SEQ ID NO:1; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:2. The amino acid sequence set forth in SEQ ID NO:2 was evaluated by BLASTP, yielding a pLog value of 210.70 versus the mouse SUG1 sequence. The sequence of the entire maize contig containing clones cs1.pk0051.b7, ctaln.pk0056.c11 and cr1n.pk0096.b2 was determined and is shown in SEQ ID NO:5; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:6. The amino acid sequence set forth in SEQ ID NO:6 was evaluated by BLASTP, yielding a pLog value of 217.03 versus the mouse SUG1 sequence. Figure 1 presents an alignment of the amino acid sequences set forth in SEQ ID NOs:2 the mouse SUG1 sequence. Figure 2 presents an alignment of the amino acid sequence set forth in SEQ ID NO:6 and the mouse SUG1 protein. The data in Table 2 represents a calculation of the percent identity of the amino acid sequences set forth in SEQ ID NOs:2 and 6 and the mouse SUG1 sequence.

TABLE 2
Percent Identity of Amino Acid Sequences Deduced From the Nucleotide Sequences of cDNA Clones Encoding Polypeptides Homologous to SUG1

Clone	SEQ ID NO.	Percent Identity to Z54219
se1.pk0023.b5	2	80
Contig composed of:	6	81
cs1.pk0051.b7		
ctaln.pk0056.c11		
cr1n.pk0096.b2		

Sequence alignments and percent identity calculations were performed by the Jotun Hein method using the Megalign program of DNASTar™ sequence analysis software (DNASTAR Inc. 1228 South Park Street, Madison Wisconsin, 53715).

- 5 Sequence alignments and BLAST scores and probabilities indicate that the instant nucleic acid fragments encode entire or nearly entire SUG1 protein. These sequences represent the first plant sequences encoding a SUG1 protein.

EXAMPLE 4

Expression of Chimeric Genes in Monocot Cells

- A chimeric gene comprising a cDNA encoding a plant SUG1 homolog in sense orientation with respect to the maize 27 kD zein promoter that is located 5' to the cDNA fragment, and the 10 kD zein 3' end that is located 3' to the cDNA fragment, can be constructed. The cDNA fragment of this gene may be generated by polymerase chain reaction (PCR) of the cDNA clone using appropriate oligonucleotide primers. Cloning sites (NcoI or SmaI) can be incorporated into the oligonucleotides to provide proper orientation of the DNA fragment when inserted into the digested vector pML103 as described below. Amplification is then performed in a standard PCR. The amplified DNA is then digested with restriction enzymes NcoI and SmaI and fractionated on an agarose gel. The appropriate band can be isolated from the gel and combined with a 4.9 kb NcoI-SmaI fragment of the plasmid pML103. Plasmid pML103 has been deposited under the terms of the Budapest Treaty at ATCC (American Type Culture Collection, 10801 University Boulevard, Manassas, VA 20110-2209), and bears accession number ATCC 97366. The DNA segment from pML103 contains a 1.05 kb SalI-NcoI promoter fragment of the maize 27 kD zein gene and a 0.96 kb SmaI-SalI fragment from the 3' end of the maize 10 kD zein gene in the vector pGem9Zf(+) (Promega). Vector and insert DNA can be ligated at 15°C overnight, essentially as described (Maniatis). The ligated DNA may then be used to transform *E. coli* XL1-Blue (Epicurian Coli XL-1 Blue™; Stratagene). Bacterial transformants can be screened by restriction enzyme digestion of plasmid DNA and limited nucleotide sequence analysis using the dideoxy chain termination method (Sequenase™ DNA Sequencing Kit; U.S. Biochemical). The resulting plasmid construct would comprise a chimeric gene encoding, in the 5' to 3' direction, the maize 27 kD zein promoter, a cDNA fragment encoding a plant SUG1 protein, and the 10 kD zein 3' region.

- The chimeric gene described above can then be introduced into corn cells by the following procedure. Immature corn embryos can be dissected from developing caryopses derived from crosses of the inbred corn lines H99 and LH132. The embryos are isolated 10 to 11 days after pollination when they are 1.0 to 1.5 mm long. The embryos are then placed with the axis-side facing down and in contact with agarose-solidified N6 medium (Chu et al., (1975) *Sci. Sin. Peking* 18:659-668). The embryos are kept in the dark at 27°C. Friable embryogenic callus consisting of undifferentiated masses of cells with somatic proembryoids and embryoids borne on suspensor structures proliferates from the scutellum

of these immature embryos. The embryogenic callus isolated from the primary explant can be cultured on N6 medium and sub-cultured on this medium every 2 to 3 weeks.

The plasmid, p35S/Ac (obtained from Dr. Peter Eckes, Hoechst Ag, Frankfurt, Germany) may be used in transformation experiments in order to provide for a selectable marker. This plasmid contains the *Pat* gene (see European Patent Publication 0 242 236) which encodes phosphinothricin acetyl transferase (PAT). The enzyme PAT confers resistance to herbicidal glutamine synthetase inhibitors such as phosphinothricin. The *pat* gene in p35S/Ac is under the control of the 35S promoter from Cauliflower Mosaic Virus (Odell et al. (1985) *Nature* 313:810-812) and the 3' region of the nopaline synthase gene from the T-DNA of the Ti plasmid of *Agrobacterium tumefaciens*.

The particle bombardment method (Klein et al., (1987) *Nature* 327:70-73) may be used to transfer genes to the callus culture cells. According to this method, gold particles (1 μ m in diameter) are coated with DNA using the following technique. Ten μ g of plasmid DNAs are added to 50 μ L of a suspension of gold particles (60 mg per mL). Calcium chloride (50 μ L of a 2.5 M solution) and spermidine free base (20 μ L of a 1.0 M solution) are added to the particles. The suspension is vortexed during the addition of these solutions. After 10 minutes, the tubes are briefly centrifuged (5 sec at 15,000 rpm) and the supernatant removed. The particles are resuspended in 200 μ L of absolute ethanol, centrifuged again and the supernatant removed. The ethanol rinse is performed again and the particles resuspended in a final volume of 30 μ L of ethanol. An aliquot (5 μ L) of the DNA-coated gold particles can be placed in the center of a Kapton™ flying disc (Bio-Rad Labs). The particles are then accelerated into the corn tissue with a Biolistic™ PDS-1000/He (Bio-Rad Instruments, Hercules CA), using a helium pressure of 1000 psi, a gap distance of 0.5 cm and a flying distance of 1.0 cm.

For bombardment, the embryogenic tissue is placed on filter paper over agarose-solidified N6 medium. The tissue is arranged as a thin lawn and covered a circular area of about 5 cm in diameter. The petri dish containing the tissue can be placed in the chamber of the PDS-1000/He approximately 8 cm from the stopping screen. The air in the chamber is then evacuated to a vacuum of 28 inches of Hg. The macrocarrier is accelerated with a helium shock wave using a rupture membrane that bursts when the He pressure in the shock tube reaches 1000 psi.

Seven days after bombardment the tissue can be transferred to N6 medium that contains glufosinate (2 mg per liter) and lacks casein or proline. The tissue continues to grow slowly on this medium. After an additional 2 weeks the tissue can be transferred to fresh N6 medium containing glufosinate. After 6 weeks, areas of about 1 cm in diameter of actively growing callus can be identified on some of the plates containing the glufosinate-supplemented medium. These calli may continue to grow when sub-cultured on the selective medium.

Plants can be regenerated from the transgenic callus by first transferring clusters of tissue to N6 medium supplemented with 0.2 mg per liter of 2,4-D. After two weeks the tissue can be transferred to regeneration medium (Fromm et al., (1990) *Bio/Technology* 8:833-839).

5

EXAMPLE 5

Expression of Chimeric Genes in Dicot Cells

A seed-specific expression cassette composed of the promoter and transcription terminator from the gene encoding the β subunit of the seed storage protein phaseolin from the bean *Phaseolus vulgaris* (Doyle et al. (1986) *J. Biol. Chem.* 261:9228-9238) can be used
10 for expression of the instant plant SUG1 protein in transformed soybean. The phaseolin cassette includes about 500 nucleotides upstream (5') from the translation initiation codon and about 1650 nucleotides downstream (3') from the translation stop codon of phaseolin. Between the 5' and 3' regions are the unique restriction endonuclease sites Nco I (which includes the ATG translation initiation codon), Sma I, Kpn I and Xba I. The entire cassette
15 is flanked by Hind III sites.

A nucleic acid fragment encoding a plant SUG1 protein may be generated by polymerase chain reaction (PCR) of the instant cDNA clones using appropriate oligonucleotide primers. Cloning sites can be incorporated into the oligonucleotides to provide proper orientation of the DNA fragment when inserted into the expression vector.
20 Amplification is then performed, and the isolated fragment is inserted into a pUC18 vector carrying the seed expression cassette.

Soybean embryos may then be transformed with the expression vector comprising sequences encoding a plant SUG1 protein. To induce somatic embryos, cotyledons, 3-5 mm in length dissected from surface sterilized, immature seeds of the soybean cultivar A2872,
25 can be cultured in the light or dark at 26°C on an appropriate agar medium for 6-10 weeks. Somatic embryos which produce secondary embryos are then excised and placed into a suitable liquid medium. After repeated selection for clusters of somatic embryos which multiplied as early, globular staged embryos, the suspensions are maintained as described below.

30 Soybean embryogenic suspension cultures can be maintained in 35 mL liquid media on a rotary shaker, 150 rpm, at 26°C with florescent lights on a 16:8 hour day/night schedule. Cultures are subcultured every two weeks by inoculating approximately 35 mg of tissue into 35 mL of liquid medium.

Soybean embryogenic suspension cultures may then be transformed by the method of
35 particle gun bombardment (Kline et al. (1987) *Nature* (London) 327:70, U.S. Patent No. 4,945,050). A Du Pont Biolistic™ PDS1000/HE instrument (helium retrofit) can be used for these transformations.

A selectable marker gene which can be used to facilitate soybean transformation is a chimeric gene composed of the 35S promoter from Cauliflower Mosaic Virus (Odell et

al.(1985) *Nature* 313:810-812), the hygromycin phosphotransferase gene from plasmid pJR225 (from *E. coli*; Gritz et al.(1983) *Gene* 25:179-188) and the 3' region of the nopaline synthase gene from the T-DNA of the Ti plasmid of *Agrobacterium tumefaciens*. The seed expression cassette comprising the phaseolin 5' region, the fragment encoding the plant
5 SUG1 protein, and the phaseolin 3' region can be isolated as a restriction fragment. This fragment can then be inserted into a unique restriction site of the vector carrying the marker gene.

To 50 μ L of a 60 mg/mL 1 μ m gold particle suspension is added (in order): 5 μ L DNA (1 μ g/ μ L), 20 μ L spermidine (0.1 M), and 50 μ L CaCl_2 (2.5 M). The particle preparation is
10 then agitated for three minutes, spun in a microfuge for 10 seconds and the supernatant removed. The DNA-coated particles are then washed once in 400 μ L 70% ethanol and resuspended in 40 μ L of anhydrous ethanol. The DNA/particle suspension can be sonicated three times for one second each. Five μ L of the DNA-coated gold particles are then loaded on each macro carrier disk.

15 Approximately 300-400 mg of a two-week-old suspension culture is placed in an empty 60x15 mm petri dish and the residual liquid removed from the tissue with a pipette. For each transformation experiment, approximately 5-10 plates of tissue are normally bombarded. Membrane rupture pressure is set at 1100 psi and the chamber is evacuated to a vacuum of 28 inches mercury. The tissue is placed approximately 3.5 inches away from the
20 retaining screen and bombarded three times. Following bombardment, the tissue can be divided in half and placed back into liquid and cultured as described above.

Five to seven days post bombardment, the liquid media may be exchanged with fresh media, and eleven to twelve days post bombardment with fresh media containing 50 mg/mL hygromycin. This selective media can be refreshed weekly. Seven to eight weeks post
25 bombardment, green, transformed tissue may be observed growing from untransformed, necrotic embryogenic clusters. Isolated green tissue is removed and inoculated into individual flasks to generate new, clonally propagated, transformed embryogenic suspension cultures. Each new line may be treated as an independent transformation event. These suspensions can then be subcultured and maintained as clusters of immature embryos or
30 regenerated into whole plants by maturation and germination of individual somatic embryos.

EXAMPLE 6

Expression of Chimeric Genes in Microbial Cells

For expression of a plant SUG1 protein, in a microbial cell, a cDNA encoding the instant plant SUG1 protein can be inserted into the T7 *E. coli* expression vector pBT430.
35 This vector is a derivative of pET-3a (Rosenberg et al. (1987) *Gene* 56:125-135) which employs the bacteriophage T7 RNA polymerase/T7 promoter system. Plasmid pBT430 was constructed by first destroying the EcoR I and Hind III sites in pET-3a at their original positions. An oligonucleotide adaptor containing EcoR I and Hind III sites was inserted at the BamH I site of pET-3a. This created pET-3aM with additional unique cloning sites for

insertion of genes into the expression vector. Then, the Nde I site at the position of translation initiation was converted to an Nco I site using oligonucleotide-directed mutagenesis. The DNA sequence of pET-3aM in this region, 5'-CATATGG, was converted to 5'-CCCATGG in pBT430.

5 Plasmid DNA containing a cDNA may be appropriately digested to release a nucleic acid fragment encoding the SUG1 protein coding sequence. This fragment may then be purified on a 1% NuSieve GTG™ low melting agarose gel (FMC). Buffer and agarose contain 10 µg/ml ethidium bromide for visualization of the DNA fragment. The fragment can then be purified from the agarose gel by digestion with GELase™ (Epicentre
10 Technologies) according to the manufacturer's instructions, ethanol precipitated, dried and resuspended in 20 µL of water. Appropriate oligonucleotide adapters may be ligated to the fragment using T4 DNA ligase (New England Biolabs, Beverly, MA). The fragment containing the ligated adapters can be purified from the excess adapters using low melting agarose as described above. The vector pBT430 is digested, dephosphorylated with alkaline
15 phosphatase (NEB) and deproteinized with phenol/chloroform as described above. The prepared vector pBT430 and fragment can then be ligated at 16°C for 15 hours followed by transformation into DH5 electrocompetent cells (GIBCO BRL). Transformants can be selected on agar plates containing LB media and 100 µg/mL ampicillin. Transformants containing the gene encoding the plant SUG1 protein are then screened for the correct
20 orientation with respect to the T7 promoter by restriction enzyme analysis.

For high level expression, a plasmid clone with the cDNA insert in the correct orientation relative to the T7 promoter can be transformed into *E. coli* strain BL21(DE3) (Studier et al. (1986) *J. Mol. Biol.* 189:113-130). Cultures are grown in LB medium containing ampicillin (100 mg/L) at 25°C. At an optical density at 600 nm of approximately
25 1, IPTG (isopropylthio-β-galactoside, the inducer) can be added to a final concentration of 0.4 mM and incubation can be continued for 3 h at 25°. Cells are then harvested by centrifugation and re-suspended in 50 µL of 50 mM Tris-HCl at pH 8.0 containing 0.1 mM DTT and 0.2 mM phenyl methylsulfonyl fluoride. A small amount of 1 mm glass beads can be added and the mixture sonicated 3 times for about 5 seconds each time with a microprobe
30 sonicator. The mixture is centrifuged and the protein concentration of the supernatant determined. One µg of protein from the soluble fraction of the culture can be separated by SDS-polyacrylamide gel electrophoresis. Gels can be observed for protein bands migrating at the expected molecular weight.

EXAMPLE 7

Evaluating Compounds for Their Ability to Inhibit Plant SUG1 Protein Activity

35 The plant SUG1 homologs described herein may be produced using any number of methods known to those skilled in the art. Such methods include, but are not limited to, expression in bacteria as described in Example 5, or expression in eukaryotic cell culture, *in planta*, and using viral expression systems in suitably infected organisms or cell lines. The

instant polypeptide may be expressed either as a mature form of the protein as observed *in vivo* or as a fusion protein by covalent attachment to a variety of enzymes, proteins or affinity tags. Common fusion protein partners include glutathione S-transferase ("GST"), thioredoxin ("Trx"), maltose binding protein, and C- and/or N-terminal hexahistidine polypeptide ("His)₆"). The fusion protein may be engineered with a protease recognition site at the fusion point so that fusion partner can be separated by protease digestion to yield an intact, mature polypeptide. Examples of such proteases include thrombin, enterokinase and factor Xa. However, any protease can be used which specifically cleaves the peptide connecting the fusion protein and the plant SUG1 polypeptide.

Purification of the instant polypeptide, if desired, may utilize any number of separation technologies familiar to those skilled in the art of protein purification. Examples of such methods include, but are not limited to, homogenization, filtration, centrifugation, heat denaturation, ammonium sulfate precipitation, desalting, pH precipitation, ion exchange chromatography, hydrophobic interaction chromatography and affinity chromatography, wherein the affinity ligand represents a substrate, substrate analog or inhibitor. When the polypeptide is expressed as a fusion protein, the purification protocol may include the use of an affinity resin which is specific for the fusion protein tag attached to the expressed polypeptide or an affinity resin containing ligands which are specific for the polypeptide. For example, polypeptide may be expressed as a fusion protein coupled to the C-terminus of thioredoxin. In addition, a (His)₆ peptide may be engineered into the N-terminus of the fused thioredoxin moiety to afford additional opportunities for affinity purification. Other suitable affinity resins could be synthesized by linking the appropriate ligands to any suitable resin such as Sepharose-4B. In an alternate embodiment, a thioredoxin fusion protein may be eluted using dithiothreitol; however, elution may be accomplished using other reagents which interact to displace the thioredoxin from the resin. These reagents include β-mercaptoethanol or other reduced thiol. The eluted fusion protein may be subjected to further purification by traditional means as stated above, if desired. Proteolytic cleavage of the thioredoxin fusion protein and the plant SUG1 protein may be accomplished after the fusion protein is purified or while the protein is still bound to the ThioBond™ affinity resin or other resin.

Crude, partially purified or purified polypeptide, either alone or as a fusion protein, may be utilized in assays for the evaluation of compounds for their ability to inhibit the activity of a plant SUG1 protein. Assays may be conducted under well known experimental conditions which permit optimal activity. An example of an *in vitro* assay for transcriptional coactivation activity is described by vom Bauer et al. ((1996) *EMBO J.* 15:110-124). The skilled artisan is well aware of simple modifications that could be made to the published protocols that would afford detection of inhibitors the plant SUG1 protein.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT:
 - (A) ADDRESSEE: E. I. DUPONT DE NEMOURS AND COMPANY
 - (B) STREET: 1007 MARKET STREET
 - (C) CITY: WILMINGTON
 - (D) STATE: DELAWARE
 - (E) COUNTRY: UNITED STATES OF AMERICA
 - (F) ZIP: 19898
 - (G) TELEPHONE: 302-992-4926
 - (H) TELEFAX: 302-773-0164
 - (I) TELEX: 6717325
- (ii) TITLE OF INVENTION: PLANT SUG1 HOMOLOGS
- (iii) NUMBER OF SEQUENCES: 14
- (iv) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: DISKETTE, 3.50 INCH
 - (B) COMPUTER: IBM PC COMPATIBLE
 - (C) OPERATING SYSTEM: MICROSOFT WINDOWS 95
 - (D) SOFTWARE: MICROSOFT WORD VERSION 7.0A
- (v) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
- (vi) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: 08/893,401
 - (B) FILING DATE: JULY 11, 1997
- (vii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: MAJARIAN, WILLIAM R.
 - (B) REGISTRATION NUMBER: 41,173
 - (C) REFERENCE/DOCKET NUMBER: BB-1095-A

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 1254 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:
 (A) ORGANISM: Soybean

(ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 1..1254

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

ATG GCT CTT GTA GGA GTT GAA CTG AAG CAT GCG GCG GAG GGC GTA CCG	48
Met Ala Leu Val Gly Val Glu Leu Lys His Ala Ala Glu Gly Val Pro	
1 5 10 15	
GAG GCG AAT TGC TCC GCC AAG CCC ACC AAG CAG GGC GAG GGC CTC CGC	96
Glu Ala Asn Cys Ser Ala Lys Pro Thr Lys Gln Gly Glu Gly Leu Arg	
20 25 30	
CAC TAC TAT TCT CTC AAC ATC CAC GAG CAT CAG CTC CTT CTT CGC CAA	144
His Tyr Tyr Ser Leu Asn Ile His Glu His Gln Leu Leu Leu Arg Gln	
35 40 45	
AAG ACT CAT AAC CTC AAC CGT CTC GAG GCT CAG AGA AAC GAC CTC AAT	192
Lys Thr His Asn Leu Asn Arg Leu Glu Ala Gln Arg Asn Asp Leu Asn	
50 55 60	
TCT AGG GTG AGG ATG CTG CGC GAA GAA TTA CAG CTT CTG CAG GAA CCC	240
Ser Arg Val Arg Met Leu Arg Glu Glu Leu Gln Leu Leu Gln Glu Pro	
65 70 75 80	
GGC TCT TAT GTC GGT GAA GTT GTC AAA GTA ATG GGC AAG AAC AAA GTC	288
Gly Ser Tyr Val Gly Glu Val Val Lys Val Met Gly Lys Asn Lys Val	
85 90 95	
CTT GTC AAG GTC CAC CCA GAA GGA AAA TAT GTT GTT GAC ATT GAC AAA	336
Leu Val Lys Val His Pro Glu Gly Lys Tyr Val Val Asp Ile Asp Lys	
100 105 110	
AAT ATT GAC ATT ACA AAG ATT ACT CCA TCC ACT AGA GTT GCA CTC CGC	384
Asn Ile Asp Ile Thr Lys Ile Thr Pro Ser Thr Arg Val Ala Leu Arg	
115 120 125	
AAC GAC AGT TAT GTT CTT CAC TTA GTT CTG CCA AGT AAA GTT GAT CCA	432
Asn Asp Ser Tyr Val Leu His Leu Val Leu Pro Ser Lys Val Asp Pro	
130 135 140	
TTG GTC AAT CTG ATG AAA GTT GAG AAA GTT CCC GAT TCT ACA TAT GAC	480
Leu Val Asn Leu Met Lys Val Glu Lys Val Pro Asp Ser Thr Tyr Asp	
145 150 155 160	
ATG ATT GGT GGT TTA GAC CAG CAA ATT AAA GAA ATA AAA GAG GTC ATT	528
Met Ile Gly Gly Leu Asp Gln Gln Ile Lys Glu Ile Lys Glu Val Ile	
165 170 175	
GAG CTA CCA ATC AAA CAT CCT GAG CTG TTT GAA AGT CTT GGA ATT GCA	576
Glu Leu Pro Ile Lys His Pro Glu Leu Phe Glu Ser Leu Gly Ile Ala	
180 185 190	

CAA CCA AAG GGT GTC CTG CTC TAT GGG CCA CCT GGT ACA GGT AAA ACA Gln Pro Lys Gly Val Leu Leu Tyr Gly Pro Pro Gly Thr Gly Lys Thr 195 200 205	624
TTG TTG GCT AGG GCA GTG GCT CAT CAT ACT GAC TGT ACA TTC ATC AGG Leu Leu Ala Arg Ala Val Ala His His Thr Asp Cys Thr Phe Ile Arg 210 215 220	672
GTG TCT GGT TCT GAG TTA GTT CAG AAA TAC ATT GGA GAA GGT TCT AGA Val Ser Gly Ser Glu Leu Val Gln Lys Tyr Ile Gly Glu Gly Ser Arg 225 230 235 240	720
ATG GTC AGG GAA CTT TTT GTT ATG GCC AGG GAA CAT GCT CCA TCA ATT Met Val Arg Glu Leu Phe Val Met Ala Arg Glu His Ala Pro Ser Ile 245 250 255	768
ATC TTC ATG GAT GAA ATT GAC AGT ATT GGA TCT GCT CGG ATG GAA TCT Ile Phe Met Asp Glu Ile Asp Ser Ile Gly Ser Ala Arg Met Glu Ser 260 265 270	816
GGA AGT GGC AAC GGT GAT AGT GAG GTA CAG CGT ACT ATG CTG GAA CTT Gly Ser Gly Asn Gly Asp Ser Glu Val Gln Arg Thr Met Leu Glu Leu 275 280 285	864
CTC AAC CAG TTG GAT GGA TTT GAA GCT TCA AAT AAG ATC AAG GTT TTG Leu Asn Gln Leu Asp Gly Phe Glu Ala Ser Asn Lys Ile Lys Val Leu 290 295 300	912
ATG GCC ACC AAT CGG ATT GAT ATC CTG GAT CAA GCC CTC CTT AGA CCA Met Ala Thr Asn Arg Ile Asp Ile Leu Asp Gln Ala Leu Leu Arg Pro 305 310 315 320	960
GGA CGG ATA GAC CGG AAA ATT GAA TTT CCA ACC CCT AAT GAA GAG TCT Gly Arg Ile Asp Arg Lys Ile Glu Phe Pro Thr Pro Asn Glu Glu Ser 325 330 335	1008
CGG CTG GAT ATT TTG AAA ATC CAT TCT AGA AGA ATG AAT TTA ATG CGT Arg Leu Asp Ile Leu Lys Ile His Ser Arg Arg Met Asn Leu Met Arg 340 345 350	1056
GGC ATT GAT TTG AAG AAG ATT GCC GAG AAG ATG AAT GGA GCA TCT GGT Gly Ile Asp Leu Lys Lys Ile Ala Glu Lys Met Asn Gly Ala Ser Gly 355 360 365	1104
GCT GAA CTT AAG GCT GTT TGC ACT GAA GCT GGA ATG TTT GCT TTG AGG Ala Glu Leu Lys Ala Val Cys Thr Glu Ala Gly Met Phe Ala Leu Arg 370 375 380	1152
GAG CGG AGG GTA CAC GTG ACT CAG GAG GAT TTT GAG ATG GCC GTG GCG Glu Arg Arg Val His Val Thr Gln Glu Asp Phe Glu Met Ala Val Ala 385 390 395 400	1200
AAG GTG ATG AAA AAG GAG ACT GAA AAA AAC ATG TCA TTG CGG AAG TTG Lys Val Met Lys Lys Glu Thr Glu Lys Asn Met Ser Leu Arg Lys Leu 405 410 415	1248
TGG AAG Trp Lys	1254

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 418 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

```

Met Ala Leu Val Gly Val Glu Leu Lys His Ala Ala Glu Gly Val Pro
 1           5           10           15
Glu Ala Asn Cys Ser Ala Lys Pro Thr Lys Gln Gly Glu Gly Leu Arg
          20           25           30
His Tyr Tyr Ser Leu Asn Ile His Glu His Gln Leu Leu Leu Arg Gln
      35           40           45
Lys Thr His Asn Leu Asn Arg Leu Glu Ala Gln Arg Asn Asp Leu Asn
      50           55           60
Ser Arg Val Arg Met Leu Arg Glu Glu Leu Gln Leu Leu Gln Glu Pro
      65           70           75           80
Gly Ser Tyr Val Gly Glu Val Val Lys Val Met Gly Lys Asn Lys Val
          85           90           95
Leu Val Lys Val His Pro Glu Gly Lys Tyr Val Val Asp Ile Asp Lys
          100           105           110
Asn Ile Asp Ile Thr Lys Ile Thr Pro Ser Thr Arg Val Ala Leu Arg
      115           120           125
Asn Asp Ser Tyr Val Leu His Leu Val Leu Pro Ser Lys Val Asp Pro
      130           135           140
Leu Val Asn Leu Met Lys Val Glu Lys Val Pro Asp Ser Thr Tyr Asp
      145           150           155           160
Met Ile Gly Gly Leu Asp Gln Gln Ile Lys Glu Ile Lys Glu Val Ile
          165           170           175
Glu Leu Pro Ile Lys His Pro Glu Leu Phe Glu Ser Leu Gly Ile Ala
          180           185           190
Gln Pro Lys Gly Val Leu Leu Tyr Gly Pro Pro Gly Thr Gly Lys Thr
      195           200           205
Leu Leu Ala Arg Ala Val Ala His His Thr Asp Cys Thr Phe Ile Arg
      210           215           220
Val Ser Gly Ser Glu Leu Val Gln Lys Tyr Ile Gly Glu Gly Ser Arg
      225           230           235           240
Met Val Arg Glu Leu Phe Val Met Ala Arg Glu His Ala Pro Ser Ile
          245           250           255
Ile Phe Met Asp Glu Ile Asp Ser Ile Gly Ser Ala Arg Met Glu Ser
          260           265           270
Gly Ser Gly Asn Gly Asp Ser Glu Val Gln Arg Thr Met Leu Glu Leu
          275           280           285
Leu Asn Gln Leu Asp Gly Phe Glu Ala Ser Asn Lys Ile Lys Val Leu
      290           295           300
Met Ala Thr Asn Arg Ile Asp Ile Leu Asp Gln Ala Leu Leu Arg Pro
      305           310           315           320

```

Gly Arg Ile Asp Arg Lys Ile Glu Phe Pro Thr Pro Asn Glu Glu Ser
 325 330 335
 Arg Leu Asp Ile Leu Lys Ile His Ser Arg Arg Met Asn Leu Met Arg
 340 345 350
 Gly Ile Asp Leu Lys Lys Ile Ala Glu Lys Met Asn Gly Ala Ser Gly
 355 360 365
 Ala Glu Leu Lys Ala Val Cys Thr Glu Ala Gly Met Phe Ala Leu Arg
 370 375 380
 Glu Arg Arg Val His Val Thr Gln Glu Asp Phe Glu Met Ala Val Ala
 385 390 395 400
 Lys Val Met Lys Lys Glu Thr Glu Lys Asn Met Ser Leu Arg Lys Leu
 405 410 415

Trp Lys

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 1148 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:
 (A) ORGANISM: Maize

(ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 3..1148

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

GA GAG CAC ATC CAT GAC CTG CAG CTC CAG ATC CGG CAG AAG ACC CAT	47
Glu His Ile His Asp Leu Gln Leu Gln Ile Arg Gln Lys Thr His	
1 5 10 15	
AAC CTC AAC CGC CTC GAG GCC CAG CGC AAC GAC CTC AAC TCC CGA GTT	95
Asn Leu Asn Arg Leu Glu Ala Gln Arg Asn Asp Leu Asn Ser Arg Val	
20 25 30	
AGA ATG CTC AGG GAA GAG TTG CAG TTG CTT CAA GAG CCT GGC TCA TAT	143
Arg Met Leu Arg Glu Glu Leu Gln Leu Gln Glu Pro Gly Ser Tyr	
35 40 45	
GTT GGT GAG GTG GTG AAG GTC ATG GGG AAA TCA AAG GTT CTG GTG AAG	191
Val Gly Glu Val Val Lys Val Met Gly Lys Ser Lys Val Leu Val Lys	
50 55 60	
GTA CAT CCC GAA GGC AAA TAT GTG GTG GAT ATA GAT AAG AGC ATT GAT	239
Val His Pro Glu Gly Lys Tyr Val Val Asp Ile Asp Lys Ser Ile Asp	
65 70 75	
ATC ACT AAG ATC ACA CCT TCA ACA AGA GTT GCT CTT CGG AAT GAC AGC	287
Ile Thr Lys Ile Thr Pro Ser Thr Arg Val Ala Leu Arg Asn Asp Ser	
80 85 90 95	
TAT ATG CTC CAT CTG ATC CTA CCA AGC AAA GTT GAT CCA TTG GTC AAT	335
Tyr Met Leu His Leu Ile Leu Pro Ser Lys Val Asp Pro Leu Val Asn	
100 105 110	

CTC	ATG	AAA	GTT	GAG	AAG	GTT	CCG	GAT	TCT	ACC	TAT	GAT	ATG	ATT	GGA	383
Leu	Met	Lys	Val	Glu	Lys	Val	Pro	Asp	Ser	Thr	Tyr	Asp	Met	Ile	Gly	
			115					120					125			
GGC	CTT	GAC	CAG	CAA	ATT	AAA	GAG	ATC	AAA	GAG	GTC	ATT	GAG	CTT	CCA	431
Gly	Leu	Asp	Gln	Gln	Ile	Lys	Glu	Ile	Lys	Glu	Val	Ile	Glu	Leu	Pro	
			130					135					140			
ATC	AAA	CAT	CCG	GAA	CTG	TTT	GAG	AGC	CTT	GGA	ATT	GCG	CAA	CCA	AAG	479
Ile	Lys	His	Pro	Glu	Leu	Phe	Glu	Ser	Leu	Gly	Ile	Ala	Gln	Pro	Lys	
			145					150					155			
GGT	GTC	CTT	CTT	TAT	GGA	CCT	CCG	GGC	ACA	GGA	AAG	ACA	TTG	TTG	GCA	527
Gly	Val	Leu	Leu	Tyr	Gly	Pro	Pro	Gly	Thr	Gly	Lys	Thr	Leu	Leu	Ala	
					165						170				175	
CGT	GCG	GTT	GCT	CAT	CAC	ACT	GAC	TGC	ACC	TTC	ATC	AGG	GTG	TCT	GGT	575
Arg	Ala	Val	Ala	His	His	Thr	Asp	Cys	Thr	Phe	Ile	Arg	Val	Ser	Gly	
				180					185					190		
TCT	GAG	TTG	GTT	CAG	AAG	TAT	ATT	GGT	GAG	GGC	TCC	CGG	ATG	GTT	AGG	623
Ser	Glu	Leu	Val	Gln	Lys	Tyr	Ile	Gly	Glu	Gly	Ser	Arg	Met	Val	Arg	
			195					200					205			
GAA	CTC	TTT	GTT	ATG	GCC	AGG	GAA	CAT	GCA	CCA	TCC	ATT	ATA	TTT	ATG	671
Glu	Leu	Phe	Val	Met	Ala	Arg	Glu	His	Ala	Pro	Ser	Ile	Ile	Phe	Met	
			210					215					220			
GAT	GAA	ATT	GAC	TCT	ATC	GGA	TCT	GCT	AGA	ATG	GAG	TCT	GGA	ACT	GGC	719
Asp	Glu	Ile	Asp	Ser	Ile	Gly	Ser	Ala	Arg	Met	Glu	Ser	Gly	Thr	Gly	
			225			230						235				
AAC	GGT	GAC	AGT	GAA	GTT	CAG	CGT	ACT	ATG	CTT	GAA	CTT	CTA	AAC	CAG	767
Asn	Gly	Asp	Ser	Glu	Val	Gln	Arg	Thr	Met	Leu	Glu	Leu	Leu	Asn	Gln	
					245						250				255	
CTC	GAT	GGT	TTT	GAA	GCA	TCA	AAC	AAA	ATT	AAG	GTT	TTG	ATG	GCA	ACG	815
Leu	Asp	Gly	Phe	Glu	Ala	Ser	Asn	Lys	Ile	Lys	Val	Leu	Met	Ala	Thr	
				260					265					270		
AAC	AGA	ATA	GAC	ATT	TTG	GAT	CAA	GCC	CTT	CTG	AGG	CCT	GGC	CGC	ATA	863
Asn	Arg	Ile	Asp	Ile	Leu	Asp	Gln	Ala	Leu	Leu	Arg	Pro	Gly	Arg	Ile	
			275					280					285			
GAC	AGG	AAG	ATT	GAA	TTT	CCA	AAT	CCT	AAC	GAG	GAT	TCA	CGT	TTC	GAT	911
Asp	Arg	Lys	Ile	Glu	Phe	Pro	Asn	Pro	Asn	Glu	Asp	Ser	Arg	Phe	Asp	
			290					295				300				
ATC	TTG	AAG	ATC	CAT	TCA	AGA	AAA	ATG	AAC	TTG	ATG	CGT	GGC	ATT	GAT	959
Ile	Leu	Lys	Ile	His	Ser	Arg	Lys	Met	Asn	Leu	Met	Arg	Gly	Ile	Asp	
			305				310					315				
CTG	AAA	AAG	ATC	GCG	GAA	AAG	ATG	AAT	GGG	GCC	TCA	GGA	GCT	GAG	CTC	1007
Leu	Lys	Lys	Ile	Ala	Glu	Lys	Met	Asn	Gly	Ala	Ser	Gly	Ala	Glu	Leu	
					325					330					335	
AAG	GCC	GTC	TGC	ACA	GAG	GCT	GGA	ATG	TTT	GCT	CTT	CGT	GAG	AGA	AGG	1055
Lys	Ala	Val	Cys	Thr	Glu	Ala	Gly	Met	Phe	Ala	Leu	Arg	Glu	Arg	Arg	
				340					345					350		
GTG	CAC	GTT	ACC	CAG	GAG	GAC	TTC	GAG	ATG	GCA	GTG	GCC	AAG	GTG	ATG	1103
Val	His	Val	Thr	Gln	Glu	Asp	Phe	Glu	Met	Ala	Val	Ala	Lys	Val	Met	
			355					360					365			

AAG AAA GAC ACG GAG AAG AAC ATG TCC CTG CGC AAG CTC TGG AAG
Lys Lys Asp Thr Glu Lys Asn Met Ser Leu Arg Lys Leu Trp Lys
370 375 380

1148

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 382 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Glu 1	His	Ile	His	Asp 5	Leu	Gln	Leu	Gln	Ile 10	Arg	Gln	Lys	Thr	His 15	Asn
Leu	Asn	Arg	Leu 20	Glu	Ala	Gln	Arg	Asn 25	Asp	Leu	Asn	Ser	Arg 30	Val	Arg
Met	Leu	Arg 35	Glu	Glu	Leu	Gln	Leu 40	Leu	Gln	Glu	Pro	Gly 45	Ser	Tyr	Val
Gly	Glu 50	Val	Val	Lys	Val	Met 55	Gly	Lys	Ser	Lys	Val 60	Leu	Val	Lys	Val
His 65	Pro	Glu	Gly	Lys	Tyr 70	Val	Val	Asp	Ile	Asp 75	Lys	Ser	Ile	Asp	Ile 80
Thr	Lys	Ile	Thr	Pro 85	Ser	Thr	Arg	Val	Ala 90	Leu	Arg	Asn	Asp	Ser 95	Tyr
Met	Leu	His	Leu 100	Ile	Leu	Pro	Ser	Lys 105	Val	Asp	Pro	Leu	Val 110	Asn	Leu
Met	Lys	Val 115	Glu	Lys	Val	Pro	Asp 120	Ser	Thr	Tyr	Asp	Met 125	Ile	Gly	Gly
Leu	Asp 130	Gln	Gln	Ile	Lys	Glu 135	Ile	Lys	Glu	Val	Ile 140	Glu	Leu	Pro	Ile
Lys 145	His	Pro	Glu	Leu	Phe 150	Glu	Ser	Leu	Gly	Ile 155	Ala	Gln	Pro	Lys	Gly 160
Val	Leu	Leu	Tyr	Gly 165	Pro	Pro	Gly	Thr	Gly 170	Lys	Thr	Leu	Leu	Ala 175	Arg
Ala	Val	Ala	His 180	His	Thr	Asp	Cys	Thr 185	Phe	Ile	Arg	Val	Ser 190	Gly	Ser
Glu	Leu	Val 195	Gln	Lys	Tyr	Ile	Gly 200	Glu	Gly	Ser	Arg	Met 205	Val	Arg	Glu
Leu	Phe 210	Val	Met	Ala	Arg	Glu 215	His	Ala	Pro	Ser	Ile 220	Ile	Phe	Met	Asp
Glu 225	Ile	Asp	Ser	Ile	Gly 230	Ser	Ala	Arg	Met	Glu 235	Ser	Gly	Thr	Gly	Asn 240
Gly	Asp	Ser	Glu	Val 245	Gln	Arg	Thr	Met	Leu 250	Glu	Leu	Leu	Asn	Gln 255	Leu
Asp	Gly	Phe	Glu 260	Ala	Ser	Asn	Lys	Ile 265	Lys	Val	Leu	Met	Ala 270	Thr	Asn

Arg Ile Asp Ile Leu Asp Gln Ala Leu Leu Arg Pro Gly Arg Ile Asp
 275 280 285
 Arg Lys Ile Glu Phe Pro Asn Pro Asn Glu Asp Ser Arg Phe Asp Ile
 290 295 300
 Leu Lys Ile His Ser Arg Lys Met Asn Leu Met Arg Gly Ile Asp Leu
 305 310 315 320
 Lys Lys Ile Ala Glu Lys Met Asn Gly Ala Ser Gly Ala Glu Leu Lys
 325 330 335
 Ala Val Cys Thr Glu Ala Gly Met Phe Ala Leu Arg Glu Arg Arg Val
 340 345 350
 His Val Thr Gln Glu Asp Phe Glu Met Ala Val Ala Lys Val Met Lys
 355 360 365
 Lys Asp Thr Glu Lys Asn Met Ser Leu Arg Lys Leu Trp Lys
 370 375 380

2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 1358 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

CCCAAGCCAA CAAGCAAGAC CGCTTCTGCT TGTGATCCGA AACCTTTCTT CGAACGCAAA	60
AAGAACCCCA CCCGACGTAC CGCGAGCCGG CGATGGCGAC GGTGGCGATG GACATCTCGA	120
AGCCCACGCC GGCAGCGTCC GGTGACGAGG CAGCAGCGGC GCGAAGGGG AGGAGCGGCG	180
GCGGGGGCGA GGGGCTGCGG CAGTACTACC TGCAGCACAT CCATGACCTG CAGCTCCAGA	240
TCCGGCAGAA GACCCATAAC CTCAACCGCC TCGAGGCCCA GCGCAACGAC CTCAACTCCC	300
GAGTTAGAAT GCTCAGGGAA GAGTTGCAGT TGCTTCAAGA GCCTGGCTCA TATGTTGGTG	360
AGGTGGTGAA GGTGATGGGG AAATCAAAGG TTCTGGTGAA GGTACATCCC GAAGGCAAAT	420
ATGTGGTGGA TATAGATAAG AGCATTGATA TACTAAGAT CACACCTTCA ACAAGAGTTG	480
CTCTTCGGAA TGACAGCTAT ATGCTCCATC TGATCTTACC AAGCAAAGTT GATCCATTGG	540
TCAATCTCAT GAAAGTTGAG AAGGTTCCGG ATTCTACCTA TGATATGATT GGAGGCCTTG	600
ACCAGCAAAT TAAAGAGATC AAAGAGGTCA TTGAGCTTCC AATCAAACAT CCGGAAGTGT	660
TTGAGAGCCT TGGAATTGCG CAACCAAAGG GTGTCCTTCT TTATGGACCT CCGGGCACAG	720
GAAAGACATT GTTGGCACGT GCGGTTGCTC ATCACACTGA CTGCACCTTC ATCAGGGTGT	780
CTGGTTCTGA GTTGGTTCAG AAGTATATTG GTGAGGGCTC CCGGATGGTT AGGGAAGTCT	840
TTGTTATGGC CAGGGAACAT GCACCATCCA TTATATTTAT GGATGAAATT GACTCTATCG	900
GATCTGCTAG AATGGAGTCT GGAAGTGGCA ACGGTGACAG TGAAGTTCAG CGTACTATGC	960
TTGAAGTTCT AAACCAGCTC GATGGTTTTG AAGCATCAAA CAAAATTAAG GTTTTGATGG	1020

CAACGAACAG AATAGACATT TTGGATCAAG CCCTTCTGAG GCCTGGCCGC ATAGACAGGA 1080
 AGATTGAATT TCCAAATCCT AACGAGGATT CACGTTTCGA TATCTTGAAG ATCCATTCAA 1140
 GAAAAATGAA CTTGATGCGT GGCATTGATC TGAAAAAGAT CGCGGAAAAG ATGAATGGGG 1200
 CCTCAGGAGC TGAGCTCAAG GCCGTCTGCA CAGAGGCTGG AATGTTTGCT CTTCTGTAGA 1260
 GAAGGGTGCA CGTTACCCAG GAGGACTTCG AGATGGCAGT GGCCAAGGTG ATGAAGAAAG 1320
 ACACGGAGAA GAACATGTCC CTGCGCAAGC TCTGGAAG 1358

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 422 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: not relevant
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Met Ala Thr Val Ala Met Asp Ile Ser Lys Pro Thr Pro Ala Ala Ser
 1 5 10 15
 Gly Asp Glu Ala Ala Ala Ala Ala Lys Gly Arg Ser Gly Gly Gly Gly
 20 25 30
 Glu Gly Leu Arg Gln Tyr Tyr Leu Gln His Ile His Asp Leu Gln Leu
 35 40 45
 Gln Ile Arg Gln Lys Thr His Asn Leu Asn Arg Leu Glu Ala Gln Arg
 50 55 60
 Asn Asp Leu Asn Ser Arg Val Arg Met Leu Arg Glu Glu Leu Gln Leu
 65 70 75 80
 Leu Gln Glu Pro Gly Ser Tyr Val Gly Glu Val Val Lys Val Met Gly
 85 90 95
 Lys Ser Lys Val Leu Val Lys Val His Pro Glu Gly Lys Tyr Val Val
 100 105 110
 Asp Ile Asp Lys Ser Ile Asp Ile Thr Lys Ile Thr Pro Ser Thr Arg
 115 120 125
 Val Ala Leu Arg Asn Asp Ser Tyr Met Leu His Leu Ile Leu Pro Ser
 130 135 140
 Lys Val Asp Pro Leu Val Asn Leu Met Lys Val Glu Lys Val Pro Asp
 145 150 155 160
 Ser Thr Tyr Asp Met Ile Gly Gly Leu Asp Gln Gln Ile Lys Glu Ile
 165 170 175
 Lys Glu Val Ile Glu Leu Pro Ile Lys His Pro Glu Leu Phe Glu Ser
 180 185 190
 Leu Gly Ile Ala Gln Pro Lys Gly Val Leu Leu Tyr Gly Pro Pro Gly
 195 200 205
 Thr Gly Lys Thr Leu Leu Ala Arg Ala Val Ala His His Thr Asp Cys
 210 215 220

Thr Phe Ile Arg Val Ser Gly Ser Glu Leu Val Gln Lys Tyr Ile Gly
 225 230 235 240
 Glu Gly Ser Arg Met Val Arg Glu Leu Phe Val Met Ala Arg Glu His
 245 250 255
 Ala Pro Ser Ile Ile Phe Met Asp Glu Ile Asp Ser Ile Gly Ser Ala
 260 265 270
 Arg Met Glu Ser Gly Thr Gly Asn Gly Asp Ser Glu Val Gln Arg Thr
 275 280 285
 Met Leu Glu Leu Leu Asn Gln Leu Asp Gly Phe Glu Ala Ser Asn Lys
 290 295 300
 Ile Lys Val Leu Met Ala Thr Asn Arg Ile Asp Ile Leu Asp Gln Ala
 305 310 315 320
 Leu Leu Arg Pro Gly Arg Ile Asp Arg Lys Ile Glu Phe Pro Asn Pro
 325 330 335
 Asn Glu Asp Ser Arg Phe Asp Ile Leu Lys Ile His Ser Arg Lys Met
 340 345 350
 Asn Leu Met Arg Gly Ile Asp Leu Lys Lys Ile Ala Glu Lys Met Asn
 355 360 365
 Gly Ala Ser Gly Ala Glu Leu Lys Ala Val Cys Thr Glu Ala Gly Met
 370 375 380
 Phe Ala Leu Arg Glu Arg Arg Val His Val Thr Gln Glu Asp Phe Glu
 385 390 395 400
 Met Ala Val Ala Lys Val Met Lys Lys Asp Thr Glu Lys Asn Met Ser
 405 410 415
 Leu Arg Lys Leu Trp Lys
 420

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 406 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS:
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Mouse

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Met Ala Leu Asp Gly Pro Glu Gln Met Glu Leu Glu Glu Gly Lys Ala
 1 5 10 15
 Gly Ser Gly Leu Arg Gln Tyr Tyr Leu Ser Lys Ile Glu Glu Leu Gln
 20 25 30
 Leu Ile Val Asn Asp Lys Ser Gln Asn Leu Arg Arg Leu Gln Ala Gln
 35 40 45
 Arg Asn Glu Leu Asn Ala Lys Val Arg Leu Leu Arg Glu Glu Leu Gln
 50 55 60

Leu Leu Gln Glu Gln Gly Ser Tyr Val Gly Glu Val Val Arg Ala Met
 65 70 75 80
 Asp Lys Lys Lys Val Leu Val Lys Val His Pro Glu Gly Lys Phe Val
 85 90 95
 Val Asp Val Asp Lys Asn Ile Asp Ile Asn Asp Val Thr Pro Asn Cys
 100 105 110
 Arg Val Ala Leu Arg Asn Asp Ser Tyr Thr Leu His Lys Ile Leu Pro
 115 120 125
 Asn Lys Val Asp Pro Leu Val Ser Leu Met Met Val Glu Lys Val Pro
 130 135 140
 Asp Ser Thr Tyr Glu Met Ile Gly Gly Leu Asp Lys Gln Ile Lys Glu
 145 150 155 160
 Ile Lys Glu Val Ile Glu Leu Pro Val Lys His Pro Glu Leu Phe Glu
 165 170 175
 Ala Leu Gly Ile Ala Gln Pro Lys Gly Val Leu Leu Tyr Gly Pro Pro
 180 185 190
 Gly Thr Gly Lys Thr Leu Leu Ala Arg Ala Val Ala His His Thr Asp
 195 200 205
 Cys Thr Phe Ile Arg Val Ser Gly Ser Glu Leu Val Gln Lys Phe Ile
 210 215 220
 Gly Glu Gly Ala Arg Met Val Arg Glu Leu Phe Val Met Ala Arg Glu
 225 230 235 240
 His Ala Pro Ser Ile Ile Phe Met Asp Glu Ile Asp Ser Ile Gly Ser
 245 250 255
 Ser Arg Leu Glu Gly Gly Ser Gly Gly Asp Ser Glu Val Gln Arg Thr
 260 265 270
 Met Leu Glu Leu Leu Asn Gln Leu Asp Gly Phe Glu Ala Thr Lys Asn
 275 280 285
 Ile Lys Val Ile Met Ala Thr Asn Arg Ile Asp Ile Leu Asp Ser Ala
 290 295 300
 Leu Leu Arg Pro Gly Arg Ile Asp Arg Lys Ile Glu Phe Pro Pro Pro
 305 310 315 320
 Asn Glu Glu Ala Arg Leu Asp Ile Leu Lys Ile His Ser Arg Lys Met
 325 330 335
 Asn Leu Thr Arg Gly Ile Asn Leu Arg Lys Ile Ala Glu Leu Met Pro
 340 345 350
 Gly Ala Ser Gly Ala Glu Val Lys Gly Val Cys Thr Glu Ala Gly Met
 355 360 365
 Tyr Ala Leu Arg Glu Arg Arg Val His Val Thr Gln Glu Asp Phe Glu
 370 375 380
 Met Ala Val Ala Lys Val Met Gln Lys Asp Ser Glu Lys Asn Met Ser
 385 390 395 400
 Ile Lys Lys Leu Trp Lys
 405

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 405 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS:
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vi) ORIGINAL SOURCE:
 (A) ORGANISM: Yeast

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

```

Met Thr Ala Ala Val Thr Ser Ser Asn Ile Val Leu Glu Thr His Glu
  1          5          10          15
Ser Gly Ile Lys Pro Tyr Phe Glu Gln Lys Ile Gln Glu Thr Glu Leu
  20          25          30
Lys Ile Arg Ser Lys Thr Glu Asn Val Arg Arg Leu Glu Ala Gln Arg
  35          40          45
Asn Ala Leu Asn Asp Lys Val Arg Phe Ile Lys Asp Glu Leu Arg Leu
  50          55          60
Leu Gln Glu Pro Gly Ser Tyr Val Gly Glu Val Ile Lys Ile Val Ser
  65          70          75          80
Asp Lys Lys Val Leu Val Lys Val Gln Pro Glu Gly Lys Tyr Ile Val
  85          90          95
Asp Val Ala Lys Asp Ile Asn Val Lys Asp Leu Lys Ala Ser Gln Arg
 100          105          110
Val Cys Leu Arg Ser Asp Ser Tyr Met Leu His Lys Val Leu Glu Asn
 115          120          125
Lys Ala Asp Pro Leu Val Ser Leu Met Met Val Glu Lys Val Pro Asp
 130          135          140
Ser Thr Tyr Asp Met Val Gly Gly Leu Thr Lys Gln Ile Lys Glu Ile
 145          150          155          160
Lys Glu Val Ile Glu Leu Pro Val Lys His Pro Glu Leu Phe Glu Ser
 165          170          175
Leu Gly Ile Ala Gln Pro Lys Gly Val Ile Leu Tyr Gly Pro Pro Gly
 180          185          190
Thr Gly Lys Thr Leu Leu Ala Arg Ala Val Ala His His Thr Asp Cys
 195          200          205
Lys Phe Ile Arg Val Ser Gly Ala Glu Leu Val Gln Lys Tyr Ile Gly
 210          215          220
Glu Gly Ser Arg Met Val Arg Glu Leu Phe Val Met Ala Arg Glu His
 225          230          235          240
Ala Pro Ser Ile Ile Phe Met Asp Glu Ile Asp Ser Ile Gly Ser Thr
 245          250          255
Arg Val Glu Gly Ser Gly Gly Gly Asp Ser Glu Val Gln Arg Thr Met
 260          265          270

```

Leu Glu Leu Leu Asn Gln Leu Asp Gly Phe Glu Thr Ser Lys Asn Ile
 275 280 285
 Lys Ile Ile Met Ala Thr Asn Arg Leu Asp Ile Leu Asp Pro Ala Leu
 290 295 300
 Leu Arg Pro Gly Arg Ile Asp Arg Lys Ile Glu Phe Pro Pro Pro Ser
 305 310 315 320
 Val Ala Ala Arg Ala Glu Ile Leu Arg Ile His Ser Arg Lys Met Asn
 325 330 335
 Leu Thr Arg Gly Ile Asn Leu Arg Lys Val Ala Glu Lys Met Asn Gly
 340 345 350
 Cys Ser Gly Ala Asp Val Lys Gly Val Cys Thr Glu Ala Gly Met Tyr
 355 360 365
 Ala Leu Arg Glu Arg Arg Ile His Val Thr Gln Glu Asp Phe Glu Leu
 370 375 380
 Ala Val Gly Lys Val Met Asn Lys Asn Gln Glu Thr Ala Ile Ser Val
 385 390 395 400
 Ala Lys Leu Phe Lys
 405

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 406 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS:
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Human Tripl

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Met Ala Leu Asp Gly Pro Glu Gln Met Glu Leu Glu Glu Gly Lys Ala
 1 5 10 15
 Gly Ser Gly Leu Arg Gln Tyr Tyr Leu Ser Lys Ile Glu Glu Leu Gln
 20 25 30
 Leu Ile Val Asn Asp Lys Ser Gln Asn Leu Arg Arg Leu Gln Ala Gln
 35 40 45
 Arg Asn Glu Leu Asn Ala Lys Val Arg Leu Leu Arg Glu Glu Leu Gln
 50 55 60
 Leu Leu Gln Glu Gln Gly Ser Tyr Val Gly Glu Val Val Arg Ala Met
 65 70 75 80
 Asp Lys Lys Lys Val Leu Val Lys Val His Pro Glu Gly Lys Phe Val
 85 90 95
 Val Asp Val Asp Lys Asn Ile Asp Ile Asn Asp Val Thr Pro Asn Cys
 100 105 110
 Arg Val Ala Leu Arg Asn Asp Ser Tyr Thr Leu His Lys Ile Leu Pro
 115 120 125

Asn Lys Val Asp Pro Leu Val Ser Leu Met Met Val Glu Lys Val Pro
 130 135 140
 Asp Ser Thr Tyr Glu Met Ile Gly Gly Leu Asp Lys Gln Ile Lys Glu
 145 150 155 160
 Ile Lys Glu Val Ile Glu Leu Pro Val Lys His Pro Glu Leu Phe Glu
 165 170 175
 Ala Leu Gly Ile Ala Gln Pro Lys Gly Val Leu Leu Tyr Gly Pro Pro
 180 185 190
 Gly Thr Gly Lys Thr Leu Leu Ala Arg Ala Val Ala His His Thr Asp
 195 200 205
 Cys Thr Phe Ile Arg Val Ser Gly Ser Glu Leu Val Gln Lys Phe Ile
 210 215 220
 Gly Glu Gly Ala Arg Met Val Arg Glu Leu Phe Val Met Ala Arg Glu
 225 230 235 240
 His Ala Pro Ser Ile Ile Phe Met Asp Glu Ile Asp Ser Ile Gly Ser
 245 250 255
 Ser Arg Leu Glu Gly Gly Ser Gly Gly Ser Ser Glu Val Gln Arg Gln
 260 265 270
 Met Leu Glu Leu Leu Asn Gln Leu Asp Gly Phe Glu Ala Thr Lys Asn
 275 280 285
 Ile Lys Val Ile Met Ala Thr Asn Arg Ile Asp Met Leu Asp Ser Ala
 290 295 300
 Leu Leu Arg Pro Gly Arg Ile Asp Arg Lys Ile Glu Phe Pro Pro Pro
 305 310 315 320
 Asn Glu Glu Ala Arg Leu Asp Ile Leu Lys Ile His Ser Arg Lys Met
 325 330 335
 Asn Leu Thr Arg Gly Ile Asn Leu Arg Lys Ile Ala Glu Leu Met Pro
 340 345 350
 Gly Ala Ser Gly Ala Glu Val Lys Gly Val Cys Thr Glu Ala Gly Met
 355 360 365
 Tyr Ala Leu Arg Glu Arg Arg Val His Val Thr Gln Glu Asp Phe Glu
 370 375 380
 Met Ala Val Ala Lys Val Met Gln Lys Asp Ser Glu Lys Asn Met Ser
 385 390 395 400
 Ile Lys Lys Leu Trp Lys
 405

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 406 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS:
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

(vi) ORIGINAL SOURCE:
(A) ORGANISM: Human p45

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

```

Met Ala Leu Asp Gly Pro Glu Gln Met Glu Leu Glu Glu Gly Lys Ala
1          5          10          15
Gly Ser Gly Leu Arg Gln Tyr Tyr Leu Ser Lys Ile Glu Glu Leu Gln
20        25        30
Leu Ile Val Asn Asp Lys Ser Gln Asn Leu Arg Arg Leu Gln Ala Gln
35        40        45
Arg Asn Glu Leu Asn Ala Lys Val Arg Leu Leu Arg Arg Glu Leu Gln
50        55        60
Leu Leu Gln Glu Gln Gly Ser Tyr Val Gly Glu Val Val Arg Ala Met
65        70        75        80
Asp Lys Lys Lys Val Leu Val Lys Val His Pro Glu Gly Lys Phe Val
85        90        95
Val Asp Val Asp Lys Asn Ile Asp Ile Asn Asp Val Thr Pro Asn Cys
100       105
Arg Val Ala Leu Arg Asn Asp Ser Tyr Thr Leu His Lys Ile Leu Pro
115       120       125
Asn Lys Val Asp Pro Leu Val Ser Leu Met Met Val Glu Lys Val Pro
130       135       140
Asp Ser Thr Tyr Glu Met Ile Gly Gly Leu Asp Lys Gln Ile Lys Glu
145       150       155       160
Ile Lys Glu Val Ile Glu Leu Pro Val Lys His Pro Glu Leu Phe Glu
165       170       175
Ala Leu Gly Ile Ala Gln Pro Lys Gly Val Leu Leu Tyr Gly Pro Pro
180       185       190
Gly Thr Gly Lys Thr Leu Leu Ala Arg Ala Val Ala His His Thr Asp
195       200       205
Cys Thr Phe Ile Arg Val Ser Gly Ser Glu Leu Val Gln Lys Phe Ile
210       215       220
Gly Glu Gly Ala Arg Met Val Arg Glu Leu Phe Val Met Ala Arg Glu
225       230       235       240
His Ala Pro Ser Ile Ile Phe Met Asp Glu Ile Asp Ser Ile Gly Ser
245       250       255
Ser Arg Leu Glu Gly Gly Ser Gly Gly Asp Ser Glu Val Gln Arg Thr
260       265       270
Met Leu Glu Leu Leu Asn Gln Leu Asp Gly Phe Glu Ala Thr Lys Asn
275       280       285
Ile Lys Val Ile Met Ala Thr Asn Arg Ile Asp Ile Leu Asp Ser Ala
290       295       300
Leu Leu Arg Pro Gly Arg Ile Asp Arg Lys Ile Glu Phe Pro Pro Pro
305       310       315       320

```


[illegible]

(2) INFORMATION FOR SEO ID NO:11:

- ```
(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 548 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vii) IMMEDIATE SOURCE:
 (B) CLONE: wlln.pk0053.g3

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:
```

|            |            |            |            |            |             |     |
|------------|------------|------------|------------|------------|-------------|-----|
| CCCCGACCGA | CCTACCCTAA | TCGGCGATGG | CGACGGTGGC | GATGGACATC | TCGAAGCCCA  | 60  |
| CTCCAGTGGC | GTCCGGCGAC | GAGGCCGCGG | CGGCGGCCAA | GGGGAGGAGC | GGCGGAGGGG  | 120 |
| GCGAGGGGCT | GCGGCAGTAC | TACCTGCAGC | ACATCCACGA | CCTGCAGCTC | CAGATCCGGC  | 180 |
| ATAAGACCCA | CAACCTCAAC | CGTCTCGAGG | CCCAGCGCAA | TGACCTCAAC | TCCCAGAGTTA | 240 |
| GAATGCTCAG | GGAAGATTGC | ANTTGCTTCA | NGAGCCTGGC | TCATATGTTG | GTNAGGTGGT  | 300 |
| GAAGGCATGG | GGAAATCAAA | GGTTCCTGGG | NAAGGTAAAC | CCCGAAGGCA | AATANNTTGT  | 360 |
| TGATATAAAT | AANCCATTGA | TTCNCANGTT | NACACCTNCA | ACAANANCGC | CTTCGAANTG  | 420 |
| ATACNANNGC | CCATCTGTTT | TACCACCAAA | TTNNCATTGG | CAACCCCATG | AANTTAAAAG  | 480 |
| TCCTGATCAC | ATAAAANCTT | TGGGGCCTGC | CACCCTTNAG | AAATANNGTT | CTTGTTTCAA  | 540 |
| NAAAACGG   |            |            |            |            |             | 548 |

(2) INFORMATION FOR SEO ID NO:12:

- ```
(i) SEQUENCE CHARACTERISTICS:
    (A) LENGTH: 115 amino acids
    (B) TYPE: amino acid
    (C) STRANDEDNESS: not relevant
    (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(vii) IMMEDIATE SOURCE:
    (B) CLONE: wlln.pk0053.q3
```

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Met Ala Thr Val Ala Met Asp Ile Ser Lys Pro Thr Pro Val Ala Ser
 1 5 10 15
 Gly Asp Glu Ala Ala Ala Ala Lys Gly Arg Ser Gly Gly Gly Gly
 20 25 30
 Glu Gly Leu Arg Gln Tyr Tyr Leu Gln His Ile His Asp Leu Gln Leu
 35 40 45
 Gln Ile Arg His Lys Thr His Asn Leu Asn Arg Leu Glu Ala Gln Arg
 50 55 60
 Asn Asp Leu Asn Ser Arg Val Arg Met Leu Arg Glu Asp Xaa Xaa Leu
 65 70 75 80
 Leu Xaa Glu Pro Gly Ser Tyr Val Gly Xaa Val Val Lys Ala Trp Gly
 85 90 95
 Asn Gln Arg Phe Trp Val Lys Val Asn Pro Glu Gly Lys Xaa Xaa Val
 100 105 110
 Asp Ile Asn
 115

(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 593 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

- (vii) IMMEDIATE SOURCE:
 (B) CLONE: rlr6.pk0064.e10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

GTTTAAACGG TACAGGAAAA ACGCTACTAG CTCGGGCAGT TGCTCATCAC ACCGACTGTA 60
 CCTTTATTAA GGGTGT CAGG TTCCGAGTTG GTTCAGAAGT ATATTGGAGA GGGTTCCAGA 120
 ATGGTTCGTG AACTATTGT GATGGCCANA GAGCATGCAC CATCCATAAT CTTTATGGAT 180
 GAAATAGACT CCATTGGATC TGCTANAATG CAGTCANGAT CTGGGGGTGG TGATAGTGAG 240
 GTTCAACGCA CTATGCTTGA TCTTCTGAAT CAACTTGATG GCTTTGAAGC ATCAAACANA 300
 ATTAAGGTCT TATGGCGACA AATANGATGG ATATTTTGGA TCAAGCTCTC TGANGCCTGG 360
 TCGCATTGAT AGGAAGATGA ATTTCCAATC CGAATGAAGA TCCGCTTGAT ANTTGAAGAT 420
 CATTCAAGAA AAATGACTGA TCTGGATTGT CTGAAAAGAT GCAGAGAAAT GATGGGCNCT 480
 GGACAAACTA AGNGTCTNAC GAACAAGATT TGCCTCCAAA NAGGTATTCA CAGAGATCTA 540
 TGGGTGCAAG GTGAAAGNAC GAAGAATCCG NCACCGAATA AGANNCACCA ACC 593

(2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 126 amino acids
 (B) TYPE: amino acid

(C) STRANDEDNESS: not relevant
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(vii) IMMEDIATE SOURCE:
 (B) CLONE: rlr6.pk0064.e10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Gly	Thr	Gly	Lys	Thr	Leu	Leu	Ala	Arg	Ala	Val	Ala	His	His	Thr	Asp
1				5					10					15	
Cys	Thr	Phe	Xaa	Arg	Val	Ser	Gly	Ser	Glu	Leu	Val	Gln	Lys	Tyr	Ile
			20					25					30		
Gly	Glu	Gly	Ser	Arg	Met	Val	Arg	Glu	Leu	Phe	Val	Met	Ala	Xaa	Glu
		35					40					45			
His	Ala	Pro	Ser	Ile	Ile	Phe	Met	Asp	Glu	Ile	Asp	Ser	Ile	Gly	Ser
	50					55					60				
Ala	Xaa	Met	Gln	Ser	Xaa	Ser	Gly	Gly	Gly	Asp	Ser	Glu	Val	Gln	Arg
65					70					75					80
Thr	Met	Leu	Asp	Leu	Leu	Asn	Gln	Leu	Asp	Gly	Phe	Glu	Ala	Ser	Asn
				85					90					95	
Xaa	Ile	Lys	Val	Xaa	Met	Ala	Thr	Asn	Xaa	Met	Asp	Ile	Leu	Asp	Gln
			100					105					110		
Ala	Leu	Xaa	Xaa	Pro	Gly	Arg	Ile	Asp	Arg	Lys	Met	Asn	Phe		
		115					120					125			

CLAIMS

What is claimed is:

1. An isolated nucleic acid fragment encoding a plant SUG1 protein comprising a member selected from the group consisting of:

- 5 (a) an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NOs:2, 4, 6, 12, and 14;
- (b) an isolated nucleic acid fragment that is substantially similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NOs:2, 4, 6, 12, and 14; and
- 10 (c) an isolated nucleic acid fragment that is complementary to (a) or (b).

2. The isolated nucleic acid fragment of Claim 1 wherein the nucleotide sequence of the fragment is set forth in a member selected from the group consisting of SEQ ID NOs:1, 3, 5, 11, and 13.

15

3. A chimeric gene comprising the nucleic acid fragment of Claim 1 operably linked to suitable regulatory sequences.

4. A transformed host cell comprising the chimeric gene of Claim 3.

5. A method of altering the level of expression of a plant SUG1 protein in a host cell comprising:

20

- (a) transforming a host cell with the chimeric gene of Claim 3; and
- (b) growing the transformed host cell produced in step (a) under conditions that are suitable for expression of the chimeric gene

wherein expression of the chimeric gene results in production of altered levels of a plant SUG1 protein in the transformed host cell.

25

6. A method of obtaining a nucleic acid fragment encoding all or substantially all of the amino acid sequence encoding a plant SUG1 protein comprising:

- (a) probing a cDNA or genomic library with the nucleic acid fragment of Claim 1;
- 30 (b) identifying a DNA clone that hybridizes with the nucleic acid fragment of Claim 1;
- (c) isolating the DNA clone identified in step (b); and
- (d) sequencing the cDNA or genomic fragment that comprises the clone isolated in step (c)

35 wherein the sequenced nucleic acid fragment encodes all or substantially all of the amino acid sequence encoding a plant SUG1 protein.

7. A method of obtaining a nucleic acid fragment encoding a portion of an amino acid sequence encoding a plant SUG1 protein comprising:

- (a) synthesizing an oligonucleotide primer corresponding to a portion of the sequence set forth in any of SEQ ID NOs:1, 3, 5, 11, and 13; and
- 5 (b) amplifying a cDNA insert present in a cloning vector using the oligonucleotide primer of step (a) and a primer representing sequences of the cloning vector

wherein the amplified nucleic acid fragment encodes a portion of an amino acid sequence encoding a plant SUG1 protein.

10 8. The product of the method of Claim 6.

9. The product of the method of Claim 7.

10. A method for evaluating at least one compound for its ability to inhibit the activity of a plant SUG1 protein, the method comprising the steps of:

- 15 (a) transforming a host cell with a chimeric gene comprising a nucleic acid fragment encoding a plant SUG1 protein, operably linked to suitable regulatory sequences;
- (b) growing the transformed host cell under conditions that are suitable for expression of the chimeric gene wherein expression of the chimeric gene results in production of the SUG1 protein encoded by the operably
- 20 linked nucleic acid fragment in the transformed host cell;
- (c) optionally purifying the SUG1 protein expressed by the transformed host cell;
- (d) treating the SUG1 protein with a compound to be tested; and
- 25 (e) comparing the activity of the SUG1 protein that has been treated with a test compound to the activity of an untreated SUG1 protein,

thereby selecting compounds with potential for inhibitory activity.

254219	MALDGEQMELEEG	10	20	30	40	50	60	70	80	90	100	89
P47210	MALDGEQMELEEG											89
L38810	MALDGEQMELEEG											89
X66400	MTAAVTSSNIVLET											88
sel.pk0023.b5	MALVGVELKHAAEGVPEANCSAKPTQGEGLRHYSINIEHQILLROKTHNINRLEAQRNDLNSRVMLREELQILQEPGSSYVGEVVMGCKNKLVKV											100
cs1.pk0051.b7	E											64
254219	HPEGEFVWDVKNIDINDVTPNCRVALRNDSTYTLHKILPNKVDPVLSIMWKEVPDSTYEMIGGLDKQIKEIKEVIELPVKHPFELFALGIAQPKGVLLY	110	120	130	140	150	160	170	180	190	200	189
P47210	HPEGEFVWDVKNIDINDVTPNCRVALRNDSTYTLHKILPNKVDPVLSIMWKEVPDSTYEMIGGLDKQIKEIKEVIELPVKHPFELFALGIAQPKGVLLY											189
L38810	HPEGEFVWDVKNIDINDVTPNCRVALRNDSTYTLHKILPNKVDPVLSIMWKEVPDSTYEMIGGLDKQIKEIKEVIELPVKHPFELFALGIAQPKGVLLY											189
X66400	QPEGKYVDVAKDINVDLKASQVCLRSDSYMLHKVLENKADPLVSIWKEVPDSTYDMWGLTKQIKEIKEVIELPVKHPFELFALGIAQPKGVLLY											188
sel.pk0023.b5	HPEGEFVWDVKNIDINDVTPNCRVALRNDSTYTLHKILPNKVDPVLSIMWKEVPDSTYEMIGGLDKQIKEIKEVIELPVKHPFELFALGIAQPKGVLLY											200
cs1.pk0051.b7	HPEGEFVWDVKNIDINDVTPNCRVALRNDSTYTLHKILPNKVDPVLSIMWKEVPDSTYEMIGGLDKQIKEIKEVIELPVKHPFELFALGIAQPKGVLLY											164
254219	GPPGIGKTLARAVAHHTDCTFIRVSGSELVQKFI GEGARWVRELFVAREHAPSI IFMDEIDSIGSSRLEGGSG-GDSEVQRTIMEILINQIDGFEATKN	210	220	230	240	250	260	270	280	290	300	288
P47210	GPPGIGKTLARAVAHHTDCTFIRVSGSELVQKFI GEGARWVRELFVAREHAPSI IFMDEIDSIGSSRLEGGSG-GDSEVQRTIMEILINQIDGFEATKN											288
L38810	GPPGIGKTLARAVAHHTDCTFIRVSGSELVQKFI GEGARWVRELFVAREHAPSI IFMDEIDSIGSSRLEGGSG-GDSEVQRTIMEILINQIDGFEATKN											288
X66400	GPPGIGKTLARAVAHHTDCTFIRVSGAEIVQKFI GEGSRWVRELFVAREHAPSI IFMDEIDSIGSTRVE-GSGGGESEVQRTIMEILINQIDGFEATKN											287
sel.pk0023.b5	GPPGIGKTLARAVAHHTDCTFIRVSGSELVQKFI GEGSRWVRELFVAREHAPSI IFMDEIDSIGARMESGSGGDSEVQRTIMEILINQIDGFEATKN											300
cs1.pk0051.b7	GPPGIGKTLARAVAHHTDCTFIRVSGSELVQKFI GEGSRWVRELFVAREHAPSI IFMDEIDSIGARMESGSGGDSEVQRTIMEILINQIDGFEATKN											264

FIG. 1

430 Rec'd PCT/PTO 1 4 JAN 2000

Z54219	IKVIMATNRIDIIDISALLRPGRIDRDKIEFFPPNEEARLDILKIHSRKVALTRGINLRKIAELMPGASGAEVKGVCITEAGMYALRERRVHVVTQEDFEVAVA	388
P47210	IKVIMATNRIDIIDISALLRPGRIDRDKIEFFPPNEEARLDILKIHSRKVALTRGINLRKIAELMPGASGAEVKGVCITEAGMYALRERRVHVVTQEDFEVAVA	388
L38810	IKVIMATNRIDIMDSALLRPGRIDRDKIEFFPPNEEARLDILKIHSRKVALTRGINLRKIAELMPGASGAEVKGVCITEAGMYALRERRVHVVTQEDFEVAVA	388
X66400	IKVIMATNRIDIIDISALLRPGRIDRDKIEFFPPNEEARLDILKIHSRKVALTRGINLRKIAELMPGASGAEVKGVCITEAGMYALRERRVHVVTQEDFEVAVA	387
se1.pk0023.b5	IKVIMATNRIDIIDISALLRPGRIDRDKIEFFPPNEEARLDILKIHSRKVALTRGINLRKIAELMPGASGAEVKGVCITEAGMYALRERRVHVVTQEDFEVAVA	400
cs1.pk0051.b7	IKVIMATNRIDIIDISALLRPGRIDRDKIEFFPPNEEARLDILKIHSRKVALTRGINLRKIAELMPGASGAEVKGVCITEAGMYALRERRVHVVTQEDFEVAVA	364
Z54219	KVMQOSEKNNMSIKKLNK	406
P47210	KVMQOSEKNNMSIKKLNK	406
L38810	KVMQOSEKNNMSIKKLNK	406
X66400	KVMKQOETAISVAKLFL	405
se1.pk0023.b5	KVMKQOETAISVAKLFL	418
cs1.pk0051.b7	KVMKQOETAISVAKLFL	382

FIG.1 (CONTINUED)

430 Rec'd PCT/PTO 1 4 JAN 2008

FIGURE 2

1 MATVAMDISKPTPAASGDEAAAAAAKGRSGGGEGLRQYYLQHIHDLQLQIRQKTHNLNRL
2 MALVGVELKHAAEGVPEANCSA....KPTKQGEGLRHYYSLNIHEHQLLLRQKTHNLNRL
3 MALDGPQMELEEG.....KAGSGLRQYYLSKIEELQLIVNDKSQNLRLRL
4 MATVAMDISKPTPVASGDEAAAAAAKGRSGGGEGLRQYYLQHIHDLQLQIRHKTTHNLNRL

1 EAQRNDLNSRVRMLREELQLLQEPGSYVGEVVKVMGKSKVLVKVHPEGKYVVDIDKSIDI
2 EAQRNDLNSRVRMLREELQLLQEPGSYVGEVVKVMGKNKVLVKVHPEGKYVVDIDKNIDI
3 QAQRNELNAKVRLREELQLLQEQGSYVGEVVRAMDKKKVLVKVHPEGKFVVDVDKNIDI
4 EAQRNDLNSRVRMLREDXLLXEPGSYVGXVVKAWGNQRFWVKVNPPEGKXXVDIN.....

1 TKITPSTRVALRND SYMLHLILPSKVDPLVNLMMKVEKVPDSTYDMIGGLDQOIKEIKEVI
2 TKITPSTRVALRND SYVLHLVLP SKVDPLVNLMMKVEKVPDSTYDMIGGLDQOIKEIKEVI
3 NDVTPNCRVALRND SYTLHKILPNKVDPLVSLMMVEKVPDSTYEMIGGLDKOIKEIKEVI
4

1 ELPIKHPELFESLGIAQPKGVLLYGPPGTGKTLLARAVAHHTDCTFIRVSGSELVQKYIG
2 ELPIKHPELFESLGIAQPKGVLLYGPPGTGKTLLARAVAHHTDCTFIRVSGSELVQKYIG
3 ELPVKHPELFEALGIAQPKGVLLYGPPGTGKTLLARAVAHHTDCTFIRVSGSELVQKFIG
4

1 EGSRMVRELFVMAREHAPSIIIFMDEIDSIGSARMESGTGNGDSEVQRTMLELLNQLDGFE
2 EGSRMVRELFVMAREHAPSIIIFMDEIDSIGSARMESGSGNGDSEVQRTMLELLNQLDGFE
3 EGARMVRELFVMAREHAPSIIIFMDEIDSIGSSRLEGGSG.GDSEVQRTMLELLNQLDGFE
4

1 ASNKIKVLMATNRIDILDQALLRPGRIDRKIEFPNPNEDSRFDILKIHSRKMNLMRGIDL
2 ASNKIKVLMATNRIDILDQALLRPGRIDRKIEFPTPNEESRLDILKIHSRRMNLMRGIDL
3 ATKNIKVIMATNRIDILDSALLRPGRIDRKIEFPNPNEEARLDILKIHSRKMNLTRGINL
4

1 KKIAEKMNGASGAELKAVCTEAGMFALRERRVHVTQEDFEMAVAKVMKKDEKMNLSLRKL
2 KKIAEKMNGASGAELKAVCTEAGMFALRERRVHVTQEDFEMAVAKVMKKETEKMNLSLRKL
3 RKIAELMPGASGAEVKGVCTEAGMYALRERRVHVTQEDFEMAVAKVMQKDEKMNMSIKKL
4

1 WK
2 WK
3 WK
4 ..

SEP 24 1999

430 Rec'd PCT/PTO 14 JAN 2000

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 98/13992

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/29 C12Q1/68 G01N33/53

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N C12Q G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	VOM BAUR, E., ET AL.: "Differential ligand-dependent interactions between the AF-2 activating domain of nuclear receptors and the putative transcriptional intermediary factors mSUG1 and TIF1" THE EMBO JOURNAL, vol. 15, no. 1, 1996, pages 110-124, XP002081516 cited in the application see the whole document -& EMBL SEQUENCE ACCESSION NO. Z54219, 22 January 1996, XP002081517 see the whole document -- -/--	1,3-5,8, 9

☒ Further documents are listed in the continuation of box C.☐ Patent family members are listed in annex.

* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search

21 October 1998

Date of mailing of the international search report

02/11/1998

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,
Fax: (+31-70) 340-3016

Authorized officer

Maddox, A

INTERNATIONAL SEARCH REPORT

Original Application No

PCT/US 98/13992

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>PROMBONA, A., ET AL.: "Structure and expression of LeMA-1, a tomato protein belonging to the SEC18-PAS1-CDC48-TBP1 protein family of putative Mg2+-dependent ATPases"</p> <p>PLANT MOLECULAR BIOLOGY, vol. 27, 1995, pages 1109-1118, XP002081518 see the whole document -& EMBL SEQUENCE ACCESSION NO.X74426, 11 April 1994, XP002081519 see the whole document</p>	1,3-5,8, 9
X	<p>SUZUKA, I.: "Rice mRNA for homologue of Tat binding protein, complet cds."</p> <p>EMBL SEQUENCE ACCESSION NO. D17788, 12 October 1994, XP002081520 see the whole document -& SUZUKA, I., ET AL.: "Identification of cDNA clones for rice homologs of the human immunodeficiency virus-1 Tat binding protein and subunit 4 of human 26S protease (proteasome)"</p> <p>PLANT SCIENCE, vol. 103, 1994, pages 33-40, XP002081521 see the whole document</p>	1,8,9
X	<p>MINOBE, Y., ET AL.: "Rice cDNA partial sequence (C10064_1A)"</p> <p>EMB SEQUENCE ACCESSION NO. D21930, 27 November 1993, XP002081522 see the whole document</p>	1,8,9
X	<p>PHILIPPS, G., ET AL.: "A. thaliana transcribed sequeunce; clone FAI117; 5' end"</p> <p>EMBL SEQUENCE ACCESSION NO.Z33979, 31 May 1994, XP002081523 see the whole document</p>	1,8,9
X	<p>SASAKI, T., ET AL.: "Rice cDNA, partial sequence (S4995_2A)"</p> <p>EMBL SEQUENCE ACCESSION NO. D41961, 14 November 1994, XP002081524 see the whole document</p>	1,8,9
X	<p>DIETZ K.J.: "Solanum tuberosum POTATP1 mRNA, complete cds."</p> <p>EMBL SEQUENCE ACCESSION NO. U43398, 20 January 1996, XP002081562 see the whole document</p>	1,8,9

-/--

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 98/13992

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	MOUNKES, L.C., ET AL.: "Drosophila melanogaster DUG mRNA, complete cds" EMBL SEQUENCE ACCESSION NO. U97538, 8 July 1997, XP002081525 see the whole document ---	1,8,9
X	NACKEN, W.K.F.: "X.laevis XSUG1 mRNA" EMBL SEQUENCE ACCESSION NO. X81986, 2 March 1995, XP002081526 see the whole document ---	1,8,9
X	JOHNSTON, S.A.: "S.cerevisiae SUG1 gene" EMBL SEQUENCE ACCESSION NO. X66400, 4 June 1992, XP002081527 see the whole document -& SWAFFIELD, J.C., ET AL.: "Alterations in a yeast protein resembling HIV Tat-binding protein relieve requirement for an acidic activation domain in GAL4" NATURE, vol. 357, June 1992, pages 698-700, XP002081528 see the whole document ---	1,8,9
X	MAKINO, Y.: "Rat liver mRNA for proteasomal ATPase (SUG1), complete cds." EMBL SEQUENCE ACCESSION NO.D83521, 28 June 1996, XP002081529 see the whole document	1,8,9
X	-& MAKINO, Y., ET AL.: "Structures of the rat proteasomal ATPases: determination of highly conserved structural motifs and rules for their spacing" BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS, vol. 220, 1996, pages 1049-1054, XP002081530 see page 1050 ---	6
X	AKIYAMA, K., ET AL.: "cDNA cloning of a new putative ATPase subunit p45 of the human 26S proteasome, a homolog of yeast transcription factor Sug1p" FEBS LETTERS, vol. 361, 1995, pages 151-156, XP002081531 see the whole document ---	1,6-9

-/--

INTERNATIONAL SEARCH REPORT

onal Application No

PCT/US 98/13992

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	ENNIS, H.L., ET AL.: "Dictyostelium discoideum HIV1 TAT-binding protein homologue (TBP10) mRNA 3' end" EMBL SEQUENCE ACCESSION NO. L16579, no. 20-5-93, XP002081532 see the whole document	1,8,9
X	-& SHAW, D.R., ET AL.: "Molecular cloning and developmental regulation of Dictyostelium discoideum homologues of the human and yeast HIV1 TAT-binding protein" BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS, vol. 193, no. 3, 1993, pages 1291-1296, XP002081533 see the whole document	7
P,X	COVITZ, PA., ET AL.: "00516 MtrHE Medicago truncatula cDNA 5' similar to 26S protease regulatory subunit 8" EMBL SEQUENCE ACCESSION NO. AA660628, 14 November 1997, XP002081534 see the whole document	1,8,9
P,X	PERRY, D.J., ET AL.: "Picea mariana TAT-binding protein homolog (Sb71) mRNA partial sequence" EMBL SEQUENCE ACCESSION NO. AF051251, 25 March 1998, XP002081535 see the whole document	1,8,9